

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of:

HEATH, *et al.*

Serial No.: 08/699,716

Filed: 27 August 1996

For: RECOMBINANT F1-V PLAGUE VACCINE



Art Unit: 1645

Examiner: Duffy, Patricia Ann

Atty. Dckt: 003/029/SAP

AFFIDAVIT OF GEORGE W. ANDERSON, JR.

1. I, George W. Anderson, Jr., an inventor of the above-referenced application and resident of Smithsburg, MD, declare the following:
2. My curriculum vitae is attached.
3. Arthur M. Friedlander, David G. Heath, Susan L. Welkos and I are joint inventors of the subject matter disclosed in the above-referenced application.
4. From **[redacted date which is before 13 March 1996]** to February 1998, I conducted research and development on a plague vaccine comprising a F1-V fusion protein as an immunogen as part of the Army Plague Vaccine Group.
5. Before about **[redacted date which is before 13 March 1996]**, I obtained alhydrogel F1-V partial preparations from David G. Heath.
6. In my laboratory notebook I usually referred to the F1-V partial as "F1-V".
7. On **[redacted date which is before 13 March 1996]**, I began mouse challenge studies with the F1-V partial preparations I obtained from David G. Heath. The experimental protocol for the challenge studies is provided in my notebook #3598 on page 123. See Exhibit GA1.
8. The results dated **[redacted date which is before 13 March 1996]** for the challenge studies using *Yersinia pestis* strain CO92, which is F1⁺ strain, with mice immunized alhydrogel F1-V partial preparations and controls with are provided in my notebook #3598 on pages 125-126. See Exhibit GA2.
9. The results dated **[redacted date which is before 13 March 1996]** for the challenge studies using *Yersinia pestis* strain C12, which is F1⁻ strain, with mice immunized with alhydrogel F1-V partial preparations and controls with are provided in my notebook #3598 on pages 127-130. See Exhibit GA3.
10. On **[redacted date which is before 13 March 1996]**, I wrote in my notebook #3598 on page 132, that the data on page 131 of my notebook and the mouse challenge studies are the first direct evidence that the F1-V fusion (F1-V partial) can induce an immune response to both F1 and V protein. David G. Heath witnessed this page and the results to which it references. See Exhibit GA4.
11. Exhibit AF3 (GA5) is an excerpt of my notebook #3598.
12. On **[redacted date which is before 13 March 1996]**, I gave David G. Heath the protocol for formulating the F1-V whole vaccine preparations for mouse challenge assays. See Exhibit

DH16 (GA6).

13. In my laboratory notebook, I usually referred to the F1-V whole as "F1-WV".
14. Before about **[redacted date which is before 13 March 1996]**, I obtained alhydrogel F1-V whole preparations from David G. Heath and began conducting the mouse challenge studies. See Exhibit GA7.
15. The results of the challenge studies dated **[redacted date which is before 13 March 1996]** using *Yersinia pestis* strain C12 or CO92 with mice immunized with alhydrogel F1-V partial preparations and controls are provided in my notebook #3739 on pages 60-63. The results show that F1-V whole confer a protective immune response against both F⁺ and F⁻ *Yersinia pestis* strains. See Exhibit GA8.
16. From about **[redacted date which is before 13 March 1996]** to 27 August 1996, I conducted further experiments to determine the efficacy of the F1-V fusion proteins and to determine whether any refinements could be made, such as the following:
 - a. On **[redacted date which is before 13 March 1996]**, I conducted a mouse challenge study examining the long term efficacy of F1-V whole which is documented in my notebook #3739, page 75. See Exhibit GA9. The results dated **[redacted date which is before 13 March 1996]** are provided in my notebook #3739, page 85. See Exhibit GA10.
 - b. On **[redacted date which is before 13 March 1996]**, I conducted a mouse challenge study examining the range of Al concentration which maintains an adequate adjuvant response which is documented in my notebook #3739, page 88. See Exhibit GA11. The alhydrogel F1-V whole preparations were obtained from David G. Heath. I copied the notebook pages from David G. Heath's notebook and inserted in my notebook. See Exhibit GA12. The results dated **[redacted date which is before 13 March 1996]** are found in my notebook #3739, pages 104-107. See Exhibit GA13.
 - c. Exhibit GA14 shows experimental data from my notebook #3739 which evidence that from **[redacted date which is before 13 March 1996]** to 23 February 1996, I conducted various mouse challenge studies with F1-V whole.
 - d. On 3 April 1996, I obtained the results for ELISA assays of serum obtained from mice immunized with F1-V whole to determine if the sera still contained antibodies against F1 antigen and V antigen as evidenced in my notebook #3739, page 122. See Exhibit GA15.
 - e. On about 15 May 1996, I conducted a study with mice vaccinated with different amounts of F1-V whole protein. The mice were challenged subcutaneously and by aerosol with *Y. pestis*, C092 or C12. See Exhibit GA16.
 - f. On about 28 June 1996, I conducted a study with mice vaccinated with either F1-V whole or a mixture of F1 + V or Plague USP vaccine. The mice were challenged by aerosol with *Y. pestis* C092. See pages 134 and 137 of my notebook #3739, Exhibit GA17.
 - g. On about 5 July 1996, I conducted a study with mice vaccinated with either F1-V whole or F1 + V or Plague USP vaccine. The mice were challenged

subcutaneously and by aerosol with *Y. pestis*, C092 or C12. See pages 135-136 of my notebook #3739, Exhibit GA18.

17. For all the challenge studies referenced herein, I obtained most of the *Yersinia pestis* strains C12 and CO92 from Susan L. Welkos.
18. On 15 February 1996, I presented the work summarized in David G. Heath's Abstract 17. See Exhibit DH19 (GA19).
19. I left the Army Plague Vaccine Group on 26 February 1998 when I retired from the U.S. Army.
20. I have reviewed and analyzed the Titball patent and the three priority documents, UK 9505059, UK 9518946, and UK 9524825, and PCT/GB96/00571.
21. It is my opinion that prior to 13 March 1996, the filing date of PCT/GB96/00571, the inventors of the Titball patent had not conceived and/or reduced to practice a plague vaccine comprising purified F1 antigen fused to all or part of V antigen as nowhere do UK 9505059, UK 9518946, and UK 9524825 disclose isolating or purifying a protein comprising F1 antigen fused to all or part of V antigen from the host cell and other cellular components and/or administering a purified protein comprising F1 antigen fused to all or part of V antigen to a subject.
 - a. In fact, UK 9518946 is the first disclosure indicating a genetic vaccine or how a host organism may be transfected with DNA for F1 antigen and V antigen to result in a live vaccine, i.e. an attenuated host organism (such as Salmonella) which produces the antigen when administered to a subject.
 - b. As described in UK 9518946, the genetic vaccine or the live vaccine is administered to a subject such that the protein/antigen of interest is then produced in the subject.
 - c. UK 9518946 does not describe isolating the protein/antigen of interest from the host organism and purifying the protein/antigen of interest from other cellular components prior to administration to a subject.
 - d. The genetic vaccine or live vaccine described in UK 9518946 is not a purified protein comprising F1 antigen fused to all or part of V antigen which is isolated and purified from cells and other cellular components as claimed in the above-referenced application.
22. I have reviewed and analyzed the experiments and data of the Army Plague Vaccine Group and it is my opinion that the Army Plague Vaccine Group:
 - a. Conceived of a fusion protein comprising F1 antigen fused to part of V by at least **[redacted date which is before 13 March 1996]**.
 - b. Conceived of a fusion protein comprising F1 antigen fused to all of V by at least **[redacted date which is before 13 March 1996]**.
 - c. Conceived of and reduced to practice a purified fusion protein comprising F1 antigen fused to part of V by at least **[redacted date which is before 13 March 1996]**.
 - d. Conceived of and reduced to practice a purified fusion protein comprising F1

antigen fused to all of V by at least [redacted date which is before 13 March 1996].

- e. Conceived of and reduced to practice a vaccine against plague comprising a purified fusion protein comprising F1 antigen fused to part of V by at least [redacted date which is before 13 March 1996].
- f. Conceived of and reduced to practice a vaccine against plague comprising a purified fusion protein comprising F1 antigen fused to all of V by at least [redacted date which is before 13 March 1996].

23. I declare that all statements made herein of my own knowledge are believed to be true, and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under §1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.



George W. Anderson, Jr.

Date: 14 March 2007

CURRICULUM VITAE

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PROFESSIONAL EXPERIENCE:

- 2004-Present Senior Principal Engineer. Scientific responsibilities for the Operations and Sustainment of a Defense Threat Reduction sponsored multi-country epidemiological surveillance system and collaborative biological research program in the former Soviet Union.
- 2003-2004 Principal Advisor. Midwest Research Institute. General scientific guidance to the company and responsibilities for integrating the capabilities of various company divisions in projects. Continue participation in DTRA projects in the former Soviet Union (Kazakhstan, Uzbekistan and Republic of Georgia).
- Senior Program Manager. Department at Southern Research Institute was purchased by Midwest Research Institute. I continued program management of the work in the former Soviet Union until promoted to Principal Advisor.
- 1998-2003 Director, Medical Countermeasures Department. Southern Research Institute. Responsible for the management, direction, control and review of the departmental research and development programs. Established a Biological Safety Level-3 containment facility for vaccine potency testing of Department of Defense (DoD) Investigational New Product (IND) vaccines (VEE, WEE, EEE, Q fever, Tularemia) in Frederick, MD. Department provided the DoD with expertise in biological defense, biosecurity, biotechnology and

biosafety for DARPA and DTRA projects at former biological weapons facilities in the former Soviet Union (FSU). Some of the projects support Non-proliferation efforts. Projects in the FSU include on-site observations of laboratory work in BSL-2, -3 and BSL-4 containment laboratories. Worked as a consultant to the Department of the Army for the anthrax vaccine production facility in the United States. Conducted audits in BSL-3 laboratories. Immunized with most licensed and IND bio-defense vaccines. Biosafety consultant to U.S. laboratories. Technical spokes person on biosafety for dismantlement of a pilot production facility for biological warfare agents in the United States.

1998-1998 Principal Scientist, SRS Technologies. Perform and manage scientific and technical tasks requiring the assessment of chemical (CW) or biological warfare (BW) programs/capabilities of foreign countries for terrorist groups and the development of recommended measures to curb the proliferation of biological weapons and technologies.

1993-1998 Chief, Pathogenesis and Immunology Branch, U.S. Army Medical Research Institute of Infectious diseases (USAMRIID). Branch Chief responsible for directing research programs directed toward development and production of prophylactic and therapeutic modalities against bacterial diseases of potential biological warfare significance (e.g., anthrax, plague, Q fever, tularemia, glanders). Special project officer for multi-million dollar Good Laboratory Practices project which included facility upgrades, Responsible for the GLP BSL-3 laboratory setup, and project manager for multi-year preclinical project for a supplement to the anthrax vaccine license. Research associate on the clinical protocol for a supplement to the current anthrax vaccine license. Manage technical aspects of a contract for cGMP production of cell banks and recombinant *Bacillus anthracis* PA protein as diagnostic or vaccine component. Involved with nonproliferation activities with former Soviet weapons scientist. First US military officer invited into the BSL-3 containment laboratories at the State Research Center of Applied Microbiology; Obolensk, Russia.

1990-1993 Medical R&D Officer, Science and Technology Center-Europe, Frankfurt, Germany - Responsible for finding biotechnologies, products, and collaboration in Europe, the Middle East, Africa, and the former Soviet Union, which could shorten or negate the need for the R&D cycle in Defense Department laboratories. This work involved extensive travel to scientific conference and Institutes in these geographical areas and technical report writing.

- 1987-1990 Research Immunologist, U.S. Army Medical Research Institute of Infectious Diseases. Investigations included development of a congenic strain of rats, vaccine efficacy trials against an aerosol exposure, and pathogenesis studies on Rift Valley fever virus. These studies were carried out in a Biosafety Level 3 laboratory.
- 1983-1987 Graduate student at the Johns Hopkins University, while on active duty, US Army.
- 1977-1983 Research Scientist, U.S. Army Medical Research Institute of Infectious Diseases (USAMRIID), Fort Detrick, Frederick, Maryland - Was responsible for developing and investigating genetically define animal models for exotic viral and rickettsial diseases.

**ACADEMIC
APPOINTMENT:**

Associate Professor, Center for Disaster Preparedness, School of Medicine, University of Alabama

FIELD STUDIES:

- 1988 Member of a team of two who conducted an epidemiological sero-survey for phleboviral infections at the MRC Laboratory, The Gambia, and at the Institute Pasteur, Senegal, West Africa.

MILITARY SERVICE:

- 1977-1998 Retirement rank: LTC, U.S. Regular Army Commission, MSC
Army Management Staff College, graduate
Command and General Staff College, graduate

EDUCATION:

- Ph.D., 1988 The Johns Hopkins University, Baltimore, Maryland, Viral Immunology. Dissertation: *Viral and Host Determinants of Resistance to Rift Valley Fever in a Rat Model*
- M.S., 1977 Florida Institute of Technology, Melbourne, Florida, Biology.
- B.S., 1975 Florida Institute of Technology, Melbourne, Florida, Biology.

TEACHING EXPERIENCE:

- 1996 Mentor, Department of Defense, Science & Engineering Apprentice Program

- 1989-1990 Served as a thesis committee member for a master's level candidate at Hood College, Frederick, Maryland. Supervised the candidate's research.
- 1988-1989 Sponsored a Korean ophthalmologist for the 1988-1989 ROK/US Scientist/Engineer Exchange Program in my laboratory to develop a model to study Rift Valley fever ocular sequelae.
- 1975-1977 Teaching assistantship at Florida Institute of Technology with primary teaching responsibilities for microbiology and biochemistry laboratories.

MEMBERSHIP IN ACADEMIC AND PROFESSIONAL SOCIETIES:

- Membership: American Society for Microbiology
 Sigma XI Scientific Research Society

HONORS AND AWARDS:

Who's Who Among Students in American Universities – 1974, 1975
Four-year Army ROTC Scholarship
Four-year U.S. Army Long Term Health, Education and Training Program (Ph.D. scholarship to the Johns Hopkins University)
Blue Key National Honor Fraternity
Sigma XI Scientific Research Society
Scouting: Highest rank – Eagle, Highest honor – Order of the Arrow, Vigil Member
U.S. Army Army Commendation Medal
 Meritorious Service Award w/2 Oak leaf clusters
 Legion of Merit
Distinguished Professional Achievement Award, Florida Institute of Technology 2002
Letter of appreciation from Joint Program Office for Biological Defense for assistance to BioPort Corporation in obtaining FDA approval for Biological License Agreement for anthrax vaccine

- PATENTS:** U.S. Patent Number 5,320,069, "Small Animal Restraint Device"
 Patent Pending Recombinant F1-V Plague Vaccine, filing #08/699,716
 18 Dec 96

CONTINUING EDUCATION:

The Regulatory Process and Good Clinical Practices, Technology Management Integration, Inc., 1994

Regulatory Issues in Biotechnology, Univ. MD, 1995
Good Manufacturing Processes for Bioprocesses, Univ. MD, 1996
Quality Control and Quality Assurance of Biotechnology Products,
Univ. MD, 1996
Assay Validation, PDA, 1996
Intro.to GLPs and Auditing, International Center for Health &
Environmental Education, 1996
Writing and Evaluating Standard Operating Procedures for the
Regulatory Environment, International Quality Training, 1996
Intro. FDA Good Laboratory Practices & Documentation Principles,
International Quality Training, 1996
Good Laboratory Practices Regulations for Study Directors,
International Quality Training, 1996
Fundamentals & Concepts of Calibration & Metrology, PDA, 1996
Biotechnology GMP Facility Design, Construction and Validation,
Univ. MD, 1997
Advances in Filtration and Bioseparation Technologies, Pall Ultrafine
Filtration Company, Columbia, MD, 1997
Validation of Biotechnology Processes and Systems, Univ. MD, 1997
Fermentation Microbiology, American Type Culture Collection
Workshop, Rockville, MD, 1997
Fundamentals of D, F and Z Values, PDA, 1997
Basic Principles in Preparation of Sterile Dosage Forms, PDA, 1997
Parenteral Packaging: Rubber, Glass, Plastic and Metal Seals, PDA,
1997
Regulatory Compliance Training, Southern Research Institute, 1998
ISO 9001/Quality system Introductory Training, Southern Research
Institute, 1998
Introductory to Earned Value Seminar, Dynport, LLC Professional
Development, 1998
Positive Pressure Pneumatic BSL-4 Suite Training, State Research
Center of Virology and Biotechnology, "Vector", Russia 2000
Introduction To Aerosol Mechanics I & II, AAAR, 2000
USA-Russia Workshop on International Research Ethics; Institutional
Review Boards and Laboratory Animal Welfare, 2002
Introduction to Laboratory Ventilation and Design, American
Biological Safety Association, 2002
Plant Biosafety, American Biological Safety Association, 2002
Bechtel Safety Leadership Workshop, Bechtel National Inc, 2006
The Transport of diagnostic & Infectious Samples, American
Biological Safety Association, 2006
Biohazard Risk Assessment, American Biological Safety Association,
2006

Certifications:

Transport of diagnostic & Infectious Substances by Air (per ICAO
Technical Instructions & IATA DGR) valid until 15 Oct 2008

REVIEWER/CONSULTANT:

U.S. Army In-house laboratory independent Research (ILIR)
proposals, 1993-1998
U.S. Army Broad Agency Announcement proposals, 1993-1998
Experts Contact for database to assist Biological Arms Control Treaty
Office (BACTO), 1997-1998
Nonproliferation Programs IntraAgency Roundtable, 1997-1998
U.S.-Uzbek Collaborative Biotechnology Grants Program, 2000
Consultant for DoD at BioPort Corporation (anthrax vaccine
production facility), 2000-2002

PROFESSIONAL PUBLICATIONS:

1. **Anderson, G.W., Jr.**, and J.V. Osterman. 1980. Host defenses in experimental Rickettsialpox: Genetics of natural resistance to infection. *Infect. Immun.* 28: 132-136.
2. **Anderson, G.W., Jr.**, and J.V. Osterman. 1980. Host defenses in Rickettsialpox: Resistance of C3H mouse sublines. *Acta. Virol.* 24: 294-296.
3. Peters, C.J., and **G.W. Anderson, Jr.** 1981. Pathogenesis of Rift Valley fever, pp. 21-41. In *Contributions to Epidemiology and Statistics*, vol. 3. (N. Goldblum, T.A. Swartz, and M.A. Kingberg, eds.). S. Karger, Basel.
4. **Anderson, G.W., Jr.**, T.W. Slone, Jr., and C.J. Peters. 1987. Pathogenesis of Rift Valley fever virus (RVFV) in inbred rats. *Microb. Pathogen.* 2: 283-293.
5. **Anderson, G.W., Jr.**, and J.F. Smith 1987. Immunoelectron microscopy of Rift Valley fever viral morphogenesis in primary rat hepatocytes. *Virology.* 161: 91-100.
6. **Anderson, G.W., Jr.**, and C.J. Peters. 1988. Viral determinants of virulence for Rift Valley fever (RVF) in rats. *Microb. Pathogen.* 5: 241-250.
7. **Anderson, G.W., Jr.**, T.W. Slone, Jr., and C.J. Peters. 1988. The gerbil, *Meriones unguiculatus*. A model for Rift Valley fever viral encephalitis. *Archives Virology* 102: 187-196.
8. **Anderson, G.W., Jr.**, J.-F. Saluzzo, T.G. Ksiazek, J.F. Smith, W. Ennis, D. Thureen, C.J. Peters, and J.P. Digoutte. 1989. Comparison of in vitro and in vivo systems for propagation of Rift Valley fever virus from clinical specimens. *Res. Virol.* 140:129-138.
9. Saluzzo, J.F., **G.W. Anderson, Jr.**, L.A. Hodgson, J.P. Digoutte, and J.F. Smith. 1989. Antigenic and biological properties of Rift Valley fever virus isolated during the 1987 Mauritanian epidemic. *Res. Virol.* 140:155-164.
10. Peters, C.J., C.-T. Liu, **G.W. Anderson, Jr.**, J.C. Morrill, and P.B. Jahrling. 1989. Pathogenesis of viral hemorrhagic fevers: Rift Valley fever and Lassa fever contrasted. *Rev. Infect. Dis.* 11 (Suppl. 4): 5743-5749.
11. Saluzzo, J.F., **G.W. Anderson, Jr.**, J.F. Smith, D. Fontenille, and P. Coulanges. 1989. Biological and antigenic relationship between Rift Valley fever virus strains isolated in Egypt and Madagascar. *Trans. R. Soc. Trop. Med. Hyg.* 83:701.
12. Solow, R., K. Mereish, **G.W. Anderson, Jr.**, and J. Hewetson. 1990. Effect of microcystin-LR on cultured rat endothelial cells. *Med. Sci. Rés.* 18:241-244.

13. **Anderson, G.W., Jr.**, M.V. Slayter, W. Hall, and C.J. Peters. 1990. Pathogenesis of a phleboviral infection (Punta Toro virus) in Golden Syrian hamsters. *Arch. Virol.* 114: 203-212.
14. **Anderson, G.W., Jr.**, J.O. Lee, A.O. Anderson, N. Powell, J.A. Mangiafico, and G. Meadors. 1991. Efficacy of a Rift Valley fever virus vaccine against an aerosol infection in rats. *Vaccine.* 9: 710-714.
15. **Anderson, G.W., Jr.**, J.A. Rosebrock, A.J. Johnson, G.B. Jennings, and C.J. Peters. 1991. Infection of inbred rats with Rift Valley Fever virus: Development of a congenic resistant strain and observations on age-dependence of resistance. *Am. J. Trop. Med. Hyg.* 44(5): 475-480.
16. **Anderson, G.W., Jr.**, W.B. Lawrence, J-O Lee, and M. Young. 1991. A restraint for ophthalmic examination of unanesthetized rats. *Note. Laboratory Animal Science.* 41(3): 288-290.
17. Friedlander, A.M., S.L. Welkos, P.L. Worsham, G.P. Andrews, D.G. Heath, **G.W. Anderson, Jr.**, M.L.M. Pitt, J. Estep, and K. Davis. 1995. Relationship between virulence and Immunity as revealed in recent studies of the F1 capsule of *Yersinia pestis*. *Clinical Infectious Diseases.* 21(Suppl 2):S178-81.
18. Andrews, G.P., D.G. Heath, **G.W. Anderson, Jr.**, S.L. Welkos, and A.M. Friedlander. 1996. Fraction 1 capsular antigen (F1) purification from *Yersinia pestis* CO92 and an *Escherichia coli* recombinant strain and efficacy against lethal plague challenge. *Infect. Immun.* 64:2180-2187.
19. **Anderson, G.W. Jr.**, S.E.C. Leary, E.D. Williamson, R.W. Titball, S.L. Welkos, P.L. Worsham, and A.M. Friedlander. 1996. Recombinant V antigen protects mice against pneumonic and bubonic plague caused by F1-capsule-positive and -negative strains of *Yersinia pestis*. *Infect. Immun.* 64:4580-4585.
20. **Anderson, G.W. Jr.**, P.L. Worsham, C.R. Bolt, G.P. Andrews, S.L. Welkos, A.M. Friedlander, and J.P. Burans. 1997. Protection of mice from fatal bubonic and pneumonic plague by passive immunization with monoclonal antibodies against the F1 protein of *Yersinia pestis*. *Am. J. Trop. Med. Hyg.* 64:4580-4585.
21. Heath, D.G., **G.W. Anderson, Jr.**, S.L. Welkos, A.M. Friedlander, and J.M. Mauro. 1997. A recombinant capsular F1-V antigen fusion protein vaccine protects against experimental bubonic and pneumonic plague. *in Vaccines 97.* Cold Spring Harbor Laboratory Press, pp 197-200.
22. Pullen, J.K., **G.W. Anderson, Jr.**, S.L. Welkos, and A.M. Friedlander. 1998. Analysis of the *Yersinia pestis* V protein for the presence of linear antibody epitopes. *Infect. Immun.* 66:521-527.

23. **Anderson, G.W. Jr.**, D.G. Heath, C.R. Bolt, S.L. Welkos, and A.M. Friedlander. Short- and long-term efficacy of single-dose subunit vaccines against *Yersinia pestis* in mice. Am. J. Trop. Med. Hyg., 58(6): 793-799.
24. Ivins, B.E., M.L.M. Pitt, P.F. Fellows, J.W. Farchaus, G.E. Benner, D.M. Waag, S.F. Little, **G.W. Anderson, Jr.**, P.H. Gibbs, and A.M. Friedlander. 1998. Comparative efficacy of experimental anthrax vaccine candidates against inhalation anthrax in rhesus macaques. Vaccine, 16(11/12): 1141-1148.
25. Heath, D.G., **G.W. Anderson, Jr.**, J.M. Mauro, S.L. Welkos, G.P. Andrews, J. Adamovicz, and A.M. Friedlander. 1998. Protection against experimental bubonic and pneumonic plague by a recombinant capsular F1-V antigen fusion protein vaccine. Vaccine, 16(11/12): 1131-1137.
26. Andrews, G.P., S.T. Strachan, G.E. Benner, A.K. Sample, J.J. Adamovicz, **G.W. Anderson, Jr.**, S.L. Welkos, A.M. Friedlander. 1999. Protective efficacy of recombinant *Yersinia* outer proteins (Yops) against bubonic plague caused by encapsulated and non-encapsulated *Yersinia pestis*. Infect. Immun., 67(3): 1533-1537.
27. Lawrence W.B., **G.W. Anderson, Jr.**, J.O. Lee, and W.C. Hall. Ocular sequelae associated with Rift Valley fever virus (RVFV) infection in inbred rats (submitted).

PUBLISHED ABSTRACTS:

1. Rosebrock, J.A., **G.W. Anderson, Jr.**, H. Schellekens, and C.J. Peters. 1983. Differential interferon sensitivities of lethal and non-lethal strains of Rift Valley fever virus (RVFV) in vitro. *In vitro* 19: 286-287.
2. **Anderson, G.W., Jr.**, M.V. Slayter, and C.J. Peters. 1988. Pathogenesis of a phleboviral infection (Punto Toro virus) in golden Syrian hamsters. *Virus Supplement* 2: 40.
3. Ribas, J.L., M.D. Kanzer, **G.W. Anderson, Jr.**, J. Sesterhenn, and C.J. Peters. 1989. Rift Valley fever viral encephalitis in the gerbil. *J. Neuropathol. Exp. Neurol.* 48: 315.
4. Ribas, J.L., M.D. Kanzer, **G.W. Anderson, Jr.**, E. Perez-Rosario, and C.J. Peters. 1990. Rift Valley fever viral encephalitis in the gerbil: Ultrastructural and immunocytochemical correlation. *J. Neuropath. Exp. Neurol.* 49:348(A).

Other publications:

Technical Report for Alternate Air Collection Media, SRS Technologies, TR98-156

PRESENTATIONS:

1. **Anderson, G.W., Jr.**, and J.V. Osterman. Susceptibility of mouse strains to *Rickettsia akari*. Presented at the 79th Annual Meeting of the American Society of Microbiologists, Los Angeles, California, 1979.
2. **Anderson, G.W., Jr.**, and J.V. Osterman. Susceptibility of mice to *Rickettsia akari*. Presented at the 1st Annual Rickettsiology Conference. Port Deposit, Maryland, 1979.
3. **Anderson, G.W., Jr.**, and C.J. Peters. Effect of immunosuppression on genetically resistant LEWIS/Mai rats to Rift Valley fever virus. Presented at the Maryland-D.C. Branch of the American Society for Microbiology Meeting, Fort Detrick, Frederick, Maryland, January 1981.
4. **Anderson, G.W., Jr.**, C.J. Peters, and T.W. Slone. Pathogenesis of Rift Valley fever virus in inbred rats. Presented at the 81st Annual Meeting of the American Society for Microbiology, Dallas, Texas, March 1981. Abstracts of the Meeting, D244.
5. Peters, C.J., and **G.W. Anderson, Jr.** Pathogenesis of Rift Valley fever and other Phlebovirus infections. Presented at the 5th International Congress of Virology, Strasbourg, France, August 1981.
6. Peters, C.J., and **G.W. Anderson, Jr.** Resistance to Phleboviruses. Presented at the U.S.-Japan Cooperative Medical Science Program, Bethesda, Maryland, 9 November 1981.
7. Peters, C.J., and **G.W. Anderson, Jr.** Pathogenesis of Phlebovirus infections. Presented at the 30th Annual Meeting of the American Society of Tropical Medicine and Hygiene, San Juan, Puerto Rico, November 1981.
8. **Anderson, G.W., Jr.**, and C.J. Peters. Role of humoral immunity in Rift Valley fever infection. Presented at the 49th Conjoint Meeting on Infectious Diseases, Ontario, Canada, 25 November 1981.
9. **Anderson, G.W., Jr.**, J.A. Rosebrock, A.J. Johnson, and C.J. Peters. Age- and dose-dependent resistance of rats to Rift Valley fever virus. Presented at the 82nd Annual Meeting of the American Society for Microbiology, Atlanta, Georgia, March 1982.
10. **Anderson, G.W., Jr.**, T.W. Slone, Jr., and C.J. Peters. A model for the encephalitic form of Rift Valley fever. Presented at the 31st Annual Meeting of the American Society of Tropical Medicine and Hygiene, Cleveland, Ohio, 1982.
11. Peters, C.J., H. Schellekens, J.A. Rosebrock, and **G.W. Anderson, Jr.** Genes, macrophages, and resistance to Rift Valley fever in the rat. Presented at the First Annual Meeting of the American Society for Virology, Ithaca, New York, 1982.

12. Peters, C.J., H. Schellenkens, J.A. Rosebrock, and **G.W. Anderson, Jr.** Genetic resistance to Rift Valley fever virus: Role of macrophages and interferon. Fourth International Conference on Comparative Virology, Banff, Canada, October 1982.
13. **Anderson, G.W., Jr.**, and J.F. Smith. Rift Valley fever virus (RVFV) maturation at the plasma membrane of rat hepatocytes as revealed by immunoelectron microscopy. Presented at the 35th Annual Meeting of the American Society of Tropical Medicine and Hygiene, Denver, Colorado; 8-11 December 1986.
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35. **Anderson, G.W.** Standard Protocols of Animal-involved Experiments in the U.S.A. Presented at the Problems of Biological and Ecological Safety. Obolensk, Russia, 22-25 May 2000.
36. **Anderson, G.** Selection of Quality Systems for Biotechnology Production Facilities. Presented at the TransBioTech 2002 – International Workshop, Protvino, Russia, 19-22 March 2002.
37. **Anderson, Jr. G.W.**, Laboratory/Field Biosafety. Presented at the Veterinary and Human Brucellosis Workshop, Almaty, Kazakhstan, 19-22 July 2004.
38. **Anderson, Jr. G.W.**, Collaborative Biosafety Efforts between the Defense Threat Reduction Agency (DTRA) and Russian Institutes. Presented at the Development of International Collaboration in Infectious Disease Research Conference, Novosibirsk, Russia, 8-10 September 2004.

File: F1-V fusion last update **REDACTED**
 Protocol: B94-02
 F1-V fusion protein immunization and challenge

see P 124-133

Investigators: CPT Heath, Dr. Welkos, LTC Anderson, COL Friedlander

Background. CPT David Heath has produced and purified a recombinant F1-V fusion protein. The protein is positive by Western blot to F1 and V. Antigen dose will be based on 10 µg F1/dose + the amount of V which is fused to it. Subcutaneous injection at nape of neck.

V-antigen used in this experiment is from Mauro. Details of the V-antigen can be obtained from CPT Heath's notebook # _____ page _____.

Purpose: Immunize and challenge mice to check on immunogenicity and protection against the CO92 strain of *Y. pestis* by sc and aerosol challenge, 50 LD₅₀.

EcF1c will be endotoxin free, same F1 as used in the active immunization experiment.

Alhydrogel, 1.3%, from SuperFos. Batch # 2043, Expiration date None, _____ µg of AL/dose

Immunization Groups: 10 Swiss Webster female mice per group from Harlan Sprague Dawley

Groups 1-9, the amount of F1 will be held constant.

Subcutaneous challenge

		Strain	# Mice
Group 1	Alhydrogel alone, days 0, 30, sc	CO92	10
Group 2	alhydrogel + 10 µg F1, days 0, 30, sc	CO92	10
Group 3	alhydrogel + 10 µg F1 urea treated, days 0, 30, sc	CO92	10
Group 4	Alhydrogel + 18.5 µg F1-V fusion protein days 0, 30, sc	CO92	10

Aerosol challenge

Group 5	Alhydrogel alone, days 0, 30, sc	CO92	10
Group 6	alhydrogel + 10 µg F1, days 0, 30, sc	CO92	10
Group 7	alhydrogel + 10 µg F1 urea treated, days 0, 30, sc	CO92	10
Group 8	Alhydrogel + 18.5 µg F1-V fusion protein, days 0, 30, sc	CO92	10
Group 9	Alhydrogel + 37.0 µg F1-V fusion protein days 0, 30, sc	<u>CO92</u>	<u>10</u>
		Total	90

1F622A6174/F-V-000
 1F633B5D66/F-V-001
 1F666A563B/F-V-002
 1F684B1B13/F-V-003
 1F73143D1D/F-V-004
 1F6336192F/F-V-005
 1F617A0402/F-V-006
 1F6373612A/F-V-007
 1F642A0E45/F-V-008
1F64667324/F-V-009
 1F610F757C/F-V-010
 1F625B356F/F-V-011
 1F62584463/F-V-012
 2007432B6B/F-V-013
 1F620D6B07/F-V-014
 1F65793A49/F-V-015

REDACTED

1F61041D5F/F-V-016
1F59570130/F-V-017
1F62776325/F-V-018
1F6E2E3015/F-V-019
1F60055D1F/F-V-020
1F66337F49/F-V-021
1F6E25735B/F-V-022
1F647B4141/F-V-023
1F6835251F/F-V-024
1F767F7A72/F-V-025, Not responding replaced with 1F617C1371
1F62530428/F-V-026
1F63717815/F-V-027
1F65122842/F-V-028
1F666F1F6D/F-V-029
1F56376371/F-V-030
1F66655A3C/F-V-031
1F663A6859/F-V-032
1F64163037/F-V-033
1F72107B64/F-V-034
1F63467B3D/F-V-035
1F645B1111/F-V-036
1F61524668/F-V-037
1F6132321C/F-V-038
1F655A0E14/F-V-039
1F661E3726/F-V-040, Not responding, replaced with 1F6134004C
1F656C5937/F-V-041
1F61135D10/F-V-042
1F6655574F/F-V-043
1F664B210F/F-V-044
7F7B0A2C5B/F-V-045
1F63737516/F-V-046
1F66240C4B/F-V-047
1F5A6F0C0C/F-V-048
1F65354106/F-V-049
1F76011A50/F-V-050
1F653D7946/F-V-051
1F624D6A48/F-V-052
1F64030B6F/F-V-053
1F6E295D6D/F-V-054
1F64303B12/F-V-055
1F635D326F/F-V-056
1F6726084C/F-V-057
1F6317796E/F-V-058
1F64024437/F-V-059
1F66056C0A/F-V-060, died from anesthesia on 17FEB95 during bleeding
1F637E6917/F-V-061
1F64653F59/F-V-062
1F643F7846/F-V-063
1F6412204B/F-V-064
7F7D261230/F-V-065
1F6132735B/F-V-066
1F635C5D45/F-V-067
1F636A1103/F-V-068
1F5F542F7F/F-V-069

1F6729527F/F-V-070
 1F63280353/F-V-071
 1F600B6D09/F-V-072
 1F77037275/F-V-073
 1F615E071B/F-V-074
 1F646F0D01/F-V-075
 1F66415A60/F-V-076
 1F65566046/F-V-077
 1F64495262/F-V-078
1F6467682E/F-V-079
 200905054D/F-V-080
 1F636F2F60/F-V-081
 1F650E541A/F-V-082
 1F637B3E45/F-V-083
 1F673C5668/F-V-084
 1F647E2E51/F-V-085
 1F663B0937/F-V-086
 1F62111658/F-V-087
 1F6320124C/F-V-088
 1F635A1F05/F-V-089

CFA from Sigma Cat#F-5881 Lot#80H8808, 10 ml/bottle

IFA from Sigma Cat#F-5506 Lot#80H8812, 10 ml/bottle

Groups 10-23 will be started one week later. The amount of V will be held constant, though the V part of the fusion protein is only about half the size of the native V.

Subcutaneous challenge

		Strain	# Mice
Group 10	Alhydrogel alone, days 0, 30, sc	C12	10
Group 11	Alhydrogel days + 27 µg F1-V fusion protein day 0, 30, sc	C12	10
Group 12	CFA + 10 µg V, days 0, IFA 30, ip	C12	10
Group 13	CFA + 10 µg V urea treated, days 0, IFA 30, ip	C12	10
Group 14	CFA + 27 µg F1-V fusion protein days 0, IFA 30, ip	C12	10
Group 15	CFA + 54 µg F1-V fusion protein days 0, IFA 30, ip	C12	10
Group 16	CFA day 0, IFA day 30 alone, ip	C12	10

Aerosol challenge

Group 17	Alhydrogel alone, days 0, 30, sc	C12	10
Group 18	Alhydrogel + 27 µg F1-V fusion protein, days 0, 30, sc	C12	10
Group 19	CFA + 10 µg V, days 0, IFA 30, ip	C12	10
Group 20	CFA + 10 µg V urea treated, days 0, IFA30, ip	C12	10
Group 21	CFA + 27 µg F1-V fusion protein day 0, IFA 30, ip	C12	10
Group 22	CFA + 54 µg F1-V fusion protein day 0, IFA 30, ip	C12	10
Group 23	CFA day 0, IFA day 30 alone, ip	<u>C12</u>	<u>10</u>
		Total	140

Group 24 CFA + 27 µg F1-V fusion protein day 0, IFA 30, ip --antibody response 10
Measure titer at 14, 27,57

Chip # for Groups 10-24, Group 10 has some mice doubled

1F646A0D06/F-V-090

7F7B107C58/F-V-091 or 1F63125319, DOUBLE CHIPPED

1F6E345D62/F-V-092 OR 1F64791E66, DOUBLE CHIPPED

1F64141752/F-V-093 OR 1F61116E01, DOUBLE CHIPPED
1F65020D6D/F-V-094 OR 9F7D25797E, DOUBLE CHIPPED
1F62032B51/F-V-095 OR 1F650627AF, DOUBLE CHIPPED
1F635D4B56/F-V-096 OR 7F7D243700, DOUBLE CHIPPED
7F7B06623C/F-V-097 OR 7F7D17224D, DOUBLE CHIPPED
7F7D23252D/F-V-098
1F630A7004/F-V-099
1F6458061F/F1-VB-001
1F66294012/F1-VB-002
1F65323119/F1-VB-003
1F68597E22/F1-VB-004
1F663B023E/F1-VB-005
1F64742F5A/F1-VB-006
1F62713757/F1-VB-007
1F664A1B16/F1-VB-008
1F624C3D76/F1-VB-009
1F60466754/F1-VB-010
1F650D343B/F1-VB-011
1F640F0668/F1-VB-012
1F684A6D42/F1-VB-013
1F627B4440/F1-VB-014
1F66362421/F1-VB-015
1F64121C4F/F1-VB-016
1F647A4340/F1-VB-017
1F757C7977/F1-VB-018
1F65625644/F1-VB-019
1F65662E68/F1-VB-020
1F655A455D/F1-VB-021, found dead, 13JAN95, cause unknown
1F63624B51/F1-VB-022
1F65487440/F1-VB-023
1F61185F09/F1-VB-024
1F63296273/F1-VB-025
1F6626094C/F1-VB-026
1F622A1C39/F1-VB-027
1F6122312D/F1-VB-028
1F6329775E/F1-VB-029
1F64652276/F1-VB-030
1F73061751/F1-VB-031
1F63672C6B/F1-VB-032
1F68406158/F1-VB-033
1F6359061F/F1-VB-034
1F653E2C12/F1-VB-035
1F60524B64/F1-VB-036
1F67274A09/F1-VB-037
200F13231B/F1-VB-038
1F620B7B79/F1-VB-039
1F66121653/F1-VB-040
1F66247661/F1-VB-041
1F652F0D40/F1-VB-042
1F63103B33/F1-VB-043
1F673D0C31/F1-VB-044
1F62757218/F1-VB-045
1F63401727/F1-VB-046
1F665F5D3F/F1-VB-047

1F66585F44/F1-VB-048
1F61195611/F1-VB-049
200737366C/F1-VB-050
1F614E2012/F1-VB-051
1F64297C58/F1-VB-052
1F75463175/F1-VB-053
1F626E157C/F1-VB-054
1F650F5419/F1-VB-055
1F64075C1A/F1-VB-056
1F756A3151/F1-VB-057
1F6121124D/F1-VB-058
1F655E405E/F1-VB-059
1F65233C1D/F1-VB-060
1F683C0B32/F1-VB-061
1F615F021F/F1-VB-062
1F71772257/F1-VB-063
1F644F713D/F1-VB-064
1F67412415/F1-VB-065
1F66606C2F/F1-VB-066
1F77087969/F1-VB-067
1F637D1F62/F1-VB-068
1F61172742/F1-VB-069
1F64511E0E/F1-VB-070
1F5756496B/F1-VB-071
1F681C4617/F1-VB-072
1F68571012/F1-VB-073
1F682F6763/F1-VB-074
1F673E4577/F1-VB-075
1F62681205/F1-VB-076
1F655F1E7F/F1-VB-077
1F65523377/F1-VB-078
1F61284810/F1-VB-079
1F6152307E/F1-VB-080
1F68382A17/F1-VB-081
1F686B1876/F1-VB-082
1F60106E03/F1-VB-083
1F63242D2D/F1-VB-084
1F637A7311/F1-VB-085
1F631F0D52/F1-VB-086
1F6378473F/F1-VB-087
1F63390C39/F1-VB-088
1F66517931/F1-VB-089
1F642C272A/F1-VB-090
1F654E703E/F1-VB-091
1F64134129/F1-VB-092
1F63355772/F1-VB-093
1F621A0263/F1-VB-094
1F64361A2D/F1-VB-095
1F61774A3F/F1-VB-096
1F65276075/F1-VB-097
1F646F0905/F1-VB-098
1F624C181B/F1-VB-099
1F684E3C6F/F1-VB-100
1F64034B2F/F1-VB-101

1F65083C38/F1-VB-102
 1F65792E55/F1-VB-103
 1F6012412E/F1-VB-104
 1F68321F28/F1-VB-105
 1F6E313909/F1-VB-106
 1F614D6F44/F1-VB-107
 1F62394402/F1-VB-108
 1F627B5C28/F1-VB-109
 1F64165A0D/F1-VB-110
 1F630B4231/F1-VB-111
 1F6629450D/F1-VB-112
 1F5F630D12/F1-VB-113
 1F657C5B25/F1-VB-114
 1F6337596E/F1-VB-115
 1F66297260/F1-VB-116
 1F66524F5A/F1-VB-117
 1F6829537D/F1-VB-118
 1F64616E2E/F1-VB-119
 1F61517936/F1-VB-120
 1F65136504/F1-VB-121
 1F66002A51/F1-VB-122
 1F5F7D1075/F1-VB-123
 1F65074134/F1-VB-124
 1F71724836/F1-VB-125
 1F656C4F41/F1-VB-126
 1F60182148/F1-VB-127
 1F6866573C/F1-VB-128
 1F62780205/F1-VB-129
 1F65080173/F1-VB-130

Schedule

Groups 1-9

22NOV94 Arrival of Swiss Webster mice, female 7-8 wks, Harlan Sprague Dawley in AA-3 (Barrier)
 01DEC94 Chipped with BioMedic Data Systems transponders in AA-3
 16DEC94 1st immunization, AA-3
 13JAN95 2nd Immunization, AA-3, day 28
 17FEB95 Bleed to determine prechallenge titers, AA-3, day 63, serum#1500-1589
 24FEB95 Challenge by aerosol & sc routes, day 70
 24MAR95 Terminal bleed, day 28 pi, titrate spleens, serum # 2403-2453

Trans # 9378 - 9432

Groups 10-23

29NOV94 Arrival of Swiss Webster mice, female 7-8 wks, Harland Sprague Dawley in AA-3 (Barrier)
 13DEC94 Chipped with BioMedic Data Systems transponders in AA-3 (SGT Zimmerman, Vet Med)
 22DEC94 1st immunization, AA-3; Crow & Fritzgerald helped
 20JAN95 2nd immunization, AA-3, day 29
 24FEB95 Bleed to dermine prechallenge titers, AA-3, day 64, serum# 1628-1768
 3MAR95 Challenge by aerosol & sc route, day 71
 31MAR95 Terminal bleed, day 28 pi, titrate spleens, serum # 2759-2800

Trans # 9433-9479

Group 24

22DEC94

1st immunization, AA-3

06JAN95

Bleed, day 14, AA-3, SERUM# 1220-1229

20JAN95

Bleed, day 27, AA-3, SERUM# 1339-1348

20JAN95

2nd immunization, day 28, AA-3

24FEB95

Bleed, day 63, AA-3, SERUM# 1768-1778 and bronchial lavage #

see page 125

Exhibit GA2

REDACTED

Project: Active Immunization F1-V with Alhydrogel

Notes: # 3598

Pathogen: Yersinia pestis strain, CO92

Route: i.p.

Dose: 63 μ g

Age: REDACTED

at 7-8 wk

Vendor: Harlan Sprague Dawley

Sex: female

EDACTED

Day postinfection

Group

Alh alone

3P1

1

Alh+F1

10 μ g

3P2

2

Alh+F1

0 μ g

area

3P3

3

Alh+F1-V

8.5 μ g

3P4

4

or Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions

iscard dead animals

se scanner to check chip number of dead mice

ark number of animals alive in each cage

REDACTED		Project: Active Immunization F1-V Alhydorgel																														
Notebook #: 3598																																
Inoculum: Yersinia pestis strain CO92																																
Route: aerosol		Dose: RUN 1 = 80 LD ₅₀ RUN 2 = 104 LD ₅₀																														
Swiss Webster		Age: 1		at 7-8wk										Vendor: Harlan Sprague Dawley										Sex: female								
REDACTED	Day -	24	25	26	27	28	1	REDACTED	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	Comments/Chip #
Day postinfection	0	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29		
Group	Cage#																															
Alh alone GP5	1																														1F6134004C/F-V-040	
	2																														1F656C5937/F-V-041	
	3																														1F61135D10/F-V-042	
	4																														1F6655574F/F-V-043	
	5																														1F664B210F/F-V-044	
	6																														7F7B0A2C5B/F-V-045	
	7																														1F63737516/F-V-046	
	8																														1F66240C4B/F-V-047	
	9																														1F5A6F0C0C/F-V-048	
	10																														1F65354106/F-V-049	
Alh+F1 10 µg GP6	11																														1F76011A50/F-V-050	
	12																														1F653D7946/F-V-051	
	13																														1F624D6A48/F-V-052	
	14																														1F64030B6F/F-V-053	
	15																														1F6E295D6D/F-V-054	
	16																														1F64303B12/F-V-055	
	17																														1F635D326F/F-V-056	
	18																														1F6726084C/F-V-057	
	19																														1F6317796E/F-V-058	
	20																														1F64024437/F-V-059	
Alh+F1 10 µg urea GP7	21	Died during bleeding of 17FEB95																												1F66056C0A/F-V-060		
	22																														1F637E6917/F-V-061	
	23																														1F64653F59/F-V-062	
	24																														1F643F7846/F-V-063	
	25																														1F6412204B/F-V-064	
	26																														7F7D261230/F-V-065	
	27																														1F6132735B/F-V-066	
	28																														1F635C5D45/F-V-067	
	29																														1F636A1103/F-V-068	
	30																														1F5F542F7F/F-V-069	
Alh+F1-V 18.5 µg GP8	31																														1F6729527F/F-V-070	
	32																														1F63280353/F-V-071	
	33																														1F600B6D09/F-V-072	
	34																														1F77037275/F-V-073	
	35																														1F615E071B/F-V-074	
	36																														1F646F0D01/F-V-075	
	37																														1F66415A60/F-V-076	
	38																														1F65566046/F-V-077	
	39																														1F64495262/F-V-078	
	40																														1F6467682E/F-V-079	
Alh+F1-V 37 µg GP9	41																														200905054D/F-V-080	
	42																														1F636F2F60/F-V-081	
	43																														1F650E541A/F-V-082	
	44																														1F637B3E45/F-V-083	
	45																														1F673C5668/F-V-084	
	46																														1F647E2E51/F-V-085	
	47																														1F663B0937/F-V-086	
	48																														1F62111658/F-V-087	
	49																														1F6320124C/F-V-088	
	50																														1F635A1F05/F-V-089	

For Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions
Discard dead animals
Use scanner to check chip # of dead animals
Mark number of mice alive

Attn: LTC Anderson

See page 123

Exhibit GA3

REDACTED

Project: Active Immunization F1-V Alhydrogel/CFA

Book #: 3598

Strain: Yersinia pestis strain C12

Exposure: Dose:

Miss Webster

Age: Arrive

REDACTED

Wk

Vendor: Harlan Sprague Dawley

Sex: female

REDACTED

infection

Cage#

alone

alone

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Comments/Chip #

1F683C0B32/F1-VB-061

1F615F021F/F1-VB-062

1F71772257/F1-VB-063

1F644F713D/F1-VB-064

1F67412415/F1-VB-065

1F66606C2F/F1-VB-066

1F77087969/F1-VB-067

1F637D1F62/F1-VB-068

1F61172742/F1-VB-069

1F64511E0E/F1-VB-070

1F5756496B/F1-VB-071

1F681C4617/F1-VB-072

1F68571012/F1-VB-073

1F682F6763/F1-VB-074

1F673E4577/F1-VB-075

1F62681205/F1-VB-076

1F655F1E7F/F1-VB-077

1F65523377/F1-VB-078

1F61284810/F1-VB-079

1F6152307E/F1-VB-080

1F68382A17/F1-VB-081

1F686B1876/F1-VB-082

1F60106E03/F1-VB-083

1F63242D2D/F1-VB-084

1F637A7311/F1-VB-085

1F631F0D52/F1-VB-086

1F6378473F/F1-VB-087

1F63390C39/F1-VB-088

1F66517931/F1-VB-089

1F642C272A/F1-VB-090

1F654E703E/F1-VB-091

1F64134129/F1-VB-092

1F63355772/F1-VB-093

1F621A0263/F1-VB-094

1F64361A2D/F1-VB-095

1F61774A3F/F1-VB-096

1F65276075/F1-VB-097

1F646F0905/F1-VB-098

1F624C181B/F1-VB-099

1F684E3C6F/F1-VB-100

1F64034B2F/F1-VB-101

1F65083C38/F1-VB-102

1F65792E55/F1-VB-103

1F6012412E/F1-VB-104

1F68321F28/F1-VB-105

1F6E313909/F1-VB-106

1F614D6F44/F1-VB-107

1F62394402/F1-VB-108

1F627B5C28/F1-VB-109

1F64165A0D/F1-VB-110

1F630B4231/F1-VB-111

1F6629450D/F1-VB-112

1F5F630D12/F1-VB-113

1F657C5B25/F1-VB-114

1F6337596E/F1-VB-115

1F66297260/F1-VB-116

1F68524F5A/F1-VB-117

1F6829537D/F1-VB-118

1F64616E2E/F1-VB-119

1F61517936/F1-VB-120

Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions

of dead animals

anner to check chip # of dead animals

number of mice alive

nose out of holder

Date: REDACTED		Project: Active Immunization F1-V Alhydrogel/CFA																																	
Notebook #: 3598																																			
Inoculum: Yersinia pestis strain C12																																			
Route: aerosol		Dose: RUN 2 = 99 LQV RUN 3 = 129 LQV																																	
Miss Webster		Age: REDACTED at 7-8wk Vendor: Harlan Sprague Dawley Sex: female																																	
REDACTED	Day postinfection	0	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	1	Comments/Chip #
Group	Cage#																																		
CFA alone	111																																	1F65136504/F1-VB	
GP23	112																																	1F66002A51/F1-VB	
	113																																	1F5F7D1075/F1-VE	
	114																																	1F65074134/F1-VB	
	115																																	1F71724836/F1-VB	
	116																																	1F656C4F41/F1-VE	
	117																																	1F60182148/F1-VB	
	118																																	1F6866573C/F1-VB	
	119																																	1F62780205/F1-VB	
	120																																	1F65080173/F1-VB	

For Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions

Discard dead animals

Use scanner to check chip # of dead animals

Mark number of mice alive

See page 127

Date: REDACTED	Project: Active Immunization F1-V Alhydrogel/CFA																														
Notebook #: 3598																															
Inoculum: Yersinia pestis strain C12																															
Route: SC	Dose: REDACTED																														
Miss Webster	Age: REDACTED																														
at 7-8wk	Vendor: Harlan Sprague Dawley																														
Sex: female																															
REDACTED	Y-	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	1
Day postinfection	0	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	
Group	Cage#																														
Alh alone	GP10	<div>1F646A0D06/F-V-090</div> <div>7F7B107C58/F-V-091 or 1F6:</div> <div>1F6E345D62/F-V-092 OR 1F:</div> <div>1F64141752/F-V-093 OR 1F:</div> <div>1F65020D6D/F-V-094 OR 9F:</div> <div>1F62032B51/F-V-095 OR 1F:</div> <div>1F635D4B56/F-V-096 OR 7F:</div> <div>7F7B06623C/F-V-097 OR 7F:</div> <div>7F7D23252D/F-V-098</div> <div>1F630A7004/F-V-099</div>																													
Alh+F1-V	GP11	<div>1F6458061F/F1-VB-001</div> <div>1F66294012/F1-VB-002</div> <div>1F65323119/F1-VB-003</div> <div>1F68597E22/F1-VB-004</div> <div>1F663B023E/F1-VB-005</div> <div>1F64742F5A/F1-VB-006</div> <div>1F62713757/F1-VB-007</div> <div>1F664A1B16/F1-VB-008</div> <div>1F624C3D76/F1-VB-009</div> <div>1F60466754/F1-VB-010</div>																													
27 µg																															
CFA+V	GP12	<div>1F650D343B/F1-VB-011</div> <div>1F640F0668/F1-VB-012</div> <div>1F684A6D42/F1-VB-013</div> <div>1F627B4440/F1-VB-014</div> <div>1F66362421/F1-VB-015</div> <div>1F64121C4F/F1-VB-016</div> <div>1F647A4340/F1-VB-017</div> <div>1F757C7977/F1-VB-018</div> <div>1F65625644/F1-VB-019</div> <div>1F65662E68/F1-VB-020</div>																													
10 µg																															
CFA+V	GP13	<div>Found dead, 13JAN95, cause unknown</div> <div>1F655A455D/F1-VB-021</div> <div>1F63624B51/F1-VB-022</div> <div>1F65487440/F1-VB-023</div> <div>1F61185F09/F1-VB-024</div> <div>1F63296273/F1-VB-025</div> <div>1F6626094C/F1-VB-026</div> <div>1F622A1C39/F1-VB-027</div> <div>1F6122312D/F1-VB-028</div> <div>1F6329775E/F1-VB-029</div> <div>1F64652276/F1-VB-030</div>																													
10 µg																															
urea																															
CFA-F1-V	GP14	<div>1F73061751/F1-VB-031</div> <div>1F63672C6B/F1-VB-032</div> <div>1F68406158/F1-VB-033</div> <div>1F6359061F/F1-VB-034</div> <div>1F653E2C12/F1-VB-035</div> <div>1F60524B84/F1-VB-036</div> <div>1F67274A09/F1-VB-037</div> <div>200F13231B/F1-VB-038</div> <div>1F620B7B79/F1-VB-039</div> <div>1F66121653/F1-VB-040</div>																													
27 µg																															
CFA-F1-V	GP15	<div>1F66247661/F1-VB-041</div> <div>1F652F0D40/F1-VB-042</div> <div>1F63103B33/F1-VB-043</div> <div>1F673D0C31/F1-VB-044</div> <div>1F62757218/F1-VB-045</div> <div>1F63401727/F1-VB-046</div> <div>1F665F5D3F/F1-VB-047</div> <div>1F66585F44/F1-VB-048</div> <div>1F61195611/F1-VB-049</div> <div>200737366C/F1-VB-050</div>																													
54 µg																															

For Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions

Discard dead animals

Use scanner to check chip # of dead animals

Mark number of mice alive

ELISA, FI & VANT.

STARTING DILUTION 1:640, SERIAL 1:2 DILUTIONS

FI AND V ANTIGENS (

FI PLATE#	SERUM#	VPLATE#	FI PLATE#	SERUM#	VPLATE#
1A F	1117	16A 16B	11A	1844 1844	26A
B	1118	16B 16B	B	1845 1845	B
2A	1119	16A 17A	12A	1846 1846	27A
B	1230	16B 17B	B	1847 1847	B
3A	1231	16A 18A	13A	1848 1848	28A
B	1232	16B 18B	B	1849 1849	B
4A	1233	19A	14A	1850 1850	29A
B	1234	19B	B	1851 NEG(N.M.)	B
5A	1235	20A	15A 15A	FI+	30A
B	1236 1836(FI)	20B	B	831	B
6A	1237 1837(FI)	21A	<p>PLATE 1-3(FI):</p> <p>200µl 10 10</p> <p>100 110 110</p> <p>REMOVED BUFFER FROM COLUMNS 2-12</p> <p>ADDED 100µl EACH SAMPLE TO COLUMN 1, 100µl BUFFER TO COLUMNS 2-12, & RETITRATED SAMPLES.</p>		
B	1238 1838(FI)	21B			
7A	1836 1236(FI)	22A			
B	1837 1237(FI)	22B			
8A	1838 1238(FI)	23A			
B	1839 1839	23B			
9A	1840 1840	24A			
B	1841 1841	24B			
10A	1842 1842	25A			
B	1843 1843	25B			

SUMMARY

FI

V

1836	POOL GP10 Alh+hydrogel alone
1837	POOL GP11 Alh+27ugF1-V
1838	POOL GP12 CFA+10ugV
1839	POOL GP13 CFA+10ugV,urea
1840	POOL GP14 CFA+27ugF1-V
1841	POOL GP15 CFA+54ugF1-V
1842	POOL GP16 CFA alone
1843	POOL GP17 Alh+hydrogel alone
1844	POOL GP18 Alh+27ugF1-V
1845	POOL GP19 CFA+10ugV
1846	POOL GP20 CFA+10ugV,urea
1847	POOL GP21 CFA+27ugF1-V
1848	POOL GP22 CFA+54ugF1-V

REDACTED

Day63	active F1-V		0	0
Day63	active F1-V		40960	81920
Day63	active F1-V		0	1310720
Day63	active F1-V		0	655360
Day63	active F1-V		163840	1310720
Day63	active F1-V		163840	1310720
Day63	active F1-V		0	1280
Day63	active F1-V		0	2560
Day63	active F1-V		40960	81920
Day63	active F1-V		0	655360
Day63	active F1-V		0	1310720
Day63	active F1-V		163840	655360
Day63	active F1-V		327680	327680

Data on page 131 is the first direct evidence that the F1-V fusion protein can induce an immune response to both the F1 and V portion of the F1-V fusion protein. This is the first proof of the concept of making a fusion protein which could be used as an immunogen in a future plague vaccine. There was some non-specific reactivity toward V, but low levels compared to the F1-V treated animals.

The second part of the proof will be the protection studies started on [REDACTED] and [REDACTED] if the protection studies work and the C092 challenge already shows protection, then this fusion protein which is unique could significantly reduce the complexity of the manufacturing process. F1-V forming an aggregate, may also increase the immunogenicity of V.

[REDACTED]
 Daniel S. Heath
 Gerald P. Johnson

Summary [REDACTED]

Serum#	Group	Treatment	Bleed Date	Day Post	F1 Titer	V Titer	Change(wells)
1836	POOL GP10	Alhydrogel alone	REDACT ED	Day63	0	0	0
1837	POOL GP11	Alh+27ugF1-V		Day63	40960	81920	163840
1838	POOL GP12	CFA+10ugV		Day63	0	1310720	1310720
1839	POOL GP13	CFA+10ugV,urea		Day63	0	655360	1310720
1840	POOL GP14	CFA+27ugF1-V		Day63	163840	1310720	1310720
1841	POOL GP15	CFA+54ugF1-V		Day63	163840	1310720	1310720
1842	POOL GP16	CFA alone		Day63	0	1280	1280
1843	POOL GP17	Alhydrogel alone		Day63	0	2560	0
1844	POOL GP18	Alh+27ugF1-V		Day63	40960	81920	163840
1845	POOL GP19	CFA+10ugV		Day63	0	655360	1310720
1846	POOL GP20	CFA+10ugV,urea		Day63	0	1310720	1310720
1847	POOL GP21	CFA+27ugF1-V		Day63	163840	655360	1310720
1848	POOL GP22	CFA+54ugF1-V		Day63	327680	327680	1310720
1849	POOL GP23	CFA alone		Day63	0	10240	1280
1850	POOL GP24	CFA+27ugF1-V		Day63	163840	163840	1310720
Controls:							
F1+ Pool					40960	20480	10240
831					0	163840	655360
Normal Mouse					0	5120	0

see page 131 & 123

file - Pitt-aerosol data [REDACTED]

EXPT. #3: [REDACTED] Aerosol and sc Plague Challenge Expt. (mice)

FOR ACTIVE F1-V IMMUNIZATION CHALLENGE 3 MAR 95
 AEROSOL - SEE PAGES 127 - 130

suspension	Target Conc./ml	no. CFU/ml	Calculated inhaled no. CFU	No. LD50s
Prespray	1.75x10e10/ml	2.8 x 10e10/ml		
C092/C12				

AGI

1

2

3

2.5 x 10e8/ml

2.6 x 10e8/ml

3.4 x 10e8/ml

see page 133
 for LD50 figures

SUBCUTANEOUS -
 Target

Date: REDACTED

PI: LTC Anderson
Agent: Plague
Strain: C12

Animal Model: Mouse

C12 LD50=1.1E+05

Wt: (Ave.): 27.69

Sex: female

see page 127-130

AGI/ml	AGI	cfu/ aerosol	MV	Inhaled Dose cfu	LD50s	Strain
2.50E+08	2.50E+09	4.17E+07	0.025	1.04E+07	94.70	C12
2.60E+08	2.60E+09	4.33E+07	0.025	1.08E+07	98.48	C12
3.40E+08	3.40E+09	5.67E+07	0.025	1.42E+07	128.79	C12

Summary REDACTED

Mouse weight

27.0

32.2

23.4

28.1

25.6

25.6

29.0

30.5

30.1

35.4

27.69g average

File: Serumbook			File Update: REDACTED			F1 TITER	V TITER
#	Group	Treatment	Blood date	Bleed day	Protocol		
220	GP24	F1-V, day 14 antibody	REDACTED	DAY14	F1-V ANTIBODY ONTOGENY	81920	40960
221	GP24	F1-V, day 14 antibody	REDACTED	DAY14	F1-V ANTIBODY ONTOGENY	20480	40960
222	GP24	F1-V, day 14 antibody	REDACTED	DAY14	F1-V ANTIBODY ONTOGENY	81920	20480
223	GP24	F1-V, day 14 antibody	REDACTED	DAY14	F1-V ANTIBODY ONTOGENY	20480	10240
224	GP24	F1-V, day 14 antibody	REDACTED	DAY14	F1-V ANTIBODY ONTOGENY	40960	640
225	GP24	F1-V, day 14 antibody	REDACTED	DAY14	F1-V ANTIBODY ONTOGENY	10240	0
226	GP24	F1-V, day 14 antibody	REDACTED	DAY14	F1-V ANTIBODY ONTOGENY	40960	10240
227	GP24	F1-V, day 14 antibody	REDACTED	DAY14	F1-V ANTIBODY ONTOGENY	40960	40960
228	GP24	F1-V, day 14 antibody	REDACTED	DAY14	F1-V ANTIBODY ONTOGENY	20480	10240
229	GP24	F1-V, day 14 antibody	REDACTED	DAY14	F1-V ANTIBODY ONTOGENY	20480	20480
Geomean(of positive values only)						31042	13934

340	GP24	F1-V, day 14 antibody	REDACTED	DAY28	F1-V ANTIBODY ONTOGENY	163840	327680
341	GP24	F1-V, day 14 antibody	REDACTED	DAY28	F1-V ANTIBODY ONTOGENY	81920	327680
342	GP24	F1-V, day 14 antibody	REDACTED	DAY28	F1-V ANTIBODY ONTOGENY	81920	1280
343	GP24	F1-V, day 14 antibody	REDACTED	DAY28	F1-V ANTIBODY ONTOGENY	81920	1310720
344	GP24	F1-V, day 14 antibody	REDACTED	DAY28	F1-V ANTIBODY ONTOGENY	20480	2560
345	GP24	F1-V, day 14 antibody	REDACTED	DAY28	F1-V ANTIBODY ONTOGENY	81920	327680
346	GP24	F1-V, day 14 antibody	REDACTED	DAY28	F1-V ANTIBODY ONTOGENY	327680	655360
347	GP24	F1-V, day 14 antibody	REDACTED	DAY28	F1-V ANTIBODY ONTOGENY	81920	40960
348	GP24	F1-V, day 14 antibody	REDACTED	DAY28	F1-V ANTIBODY ONTOGENY	40960	81920
349	GP24	F1-V, day 14 antibody	REDACTED	DAY28	F1-V ANTIBODY ONTOGENY	81920	163840
Geomean(of positive values only)						81920	94101

768	GP24	CFA+27ugF1-V	REDACTED	Day63	active F1-V	327680	1310720
769	GP24	CFA+27ugF1-V	REDACTED	Day63	active F1-V	655360	655360
770	GP24	CFA+27ugF1-V	REDACTED	Day63	active F1-V	327680	655360
771	GP24	CFA+27ugF1-V	REDACTED	Day63	active F1-V	327680	1310720
772	GP24	CFA+27ugF1-V	REDACTED	Day63	active F1-V	655360	655360
773	GP24	CFA+27ugF1-V	REDACTED	Day63	active F1-V	327680	1310720
774	GP24	CFA+27ugF1-V	REDACTED	Day63	active F1-V	163840	40960
775	GP24	CFA+27ugF1-V	REDACTED	Day63	active F1-V	163840	1310720
776	GP24	CFA+27ugF1-V	REDACTED	Day63	active F1-V	655360	1310720
777	GP24	CFA+27ugF1-V	REDACTED	Day63	active F1-V	Died	

Geomean(of positive values only)

353914

707828

Summary REDACTED

File: Serumbook			File Update:		REDACTED			
Serum #	Group	Treatment	Bleed date	Bleed day	Protocol	CHIP #	MISC	V TITER
1688	GP16	CFA alone	REDACTED	Day63	active F1-V	1F614E2012/F1-VB-051		O
1689	GP16	CFA alone		Day63	active F1-V	1F64297C58/F1-VB-052		O
1690	GP16	CFA alone		Day63	active F1-V	1F75463175/F1-VB-053		O
1691	GP16	CFA alone		Day63	active F1-V	1F626E157C/F1-VB-054		O
1692	GP16	CFA alone		Day63	active F1-V	1F650F5419/F1-VB-055		O
1693	GP16	CFA alone		Day63	active F1-V	1F64075C1A/F1-VB-056		O
1694	GP16	CFA alone		Day63	active F1-V	1F756A3151/F1-VB-057		No sample
1695	GP16	CFA alone		Day63	active F1-V	1F6121124D/F1-VB-058		640
1696	GP16	CFA alone		Day63	active F1-V	1F655E405E/F1-VB-059		O
1697	GP16	CFA alone		Day63	active F1-V	1F65233C1D/F1-VB-060		O
Geomean(of positive values only)								640

1758	GP23 CFA alone	REDACTED	Day63	active F1-V	1F65136504/F1-VB-121	1280
1759	GP23 CFA alone		Day63	active F1-V	1F66002A51/F1-VB-122	O
1760	GP23 CFA alone		Day63	active F1-V	1F5F7D1075/F1-VB-123	20480
1761	GP23 CFA alone		Day63	active F1-V	1F65074134/F1-VB-124	O
1762	GP23 CFA alone		Day63	active F1-V	1F71724836/F1-VB-125	640
1763	GP23 CFA alone		Day63	active F1-V	1F656C4F41/F1-VB-126	O
1764	GP23 CFA alone		Day63	active F1-V	1F60182148/F1-VB-127	O
1765	GP23 CFA alone		Day63	active F1-V	1F6866573C/F1-VB-128	O
1766	GP23 CFA alone		Day63	active F1-V	1F62780205/F1-VB-129	1280
1767	GP23 CFA alone		Day63	active F1-V	1F65080173/F1-VB-130	O
Geomean(of positive values only!)						2153

Controls:

Normal Mouse	0
F1+ Pool	5120
Serum 831	327680

CFA appear to increase background problem. #1760 is so high it appears to be an error of some sort. Most probably, a tube contamination or mislabeled tube.

Summary REDACTED

File: Serumbook			File Update: REDACTED				
Serum #	Group	Treatment	Bleed date	Bleed day	Protocol	F1 TITER	V TITER
2081	GP1 Pool	Alhydrogel only	REDACTED	Day63	ALH-F1-V fusion	O	O
2082	GP2 Pool	ALH+10µgF1		Day63	ALH-F1-V fusion	81920	O
2083	GP3 Pool	ALH+10µgF2urea		Day63	ALH-F1-V fusion	81920	O
2084	GP4 Pool	ALH+18.5µg F1-V		Day63	ALH-F1-V fusion	81920	163840
2085	GP5 Pool	Alhydrogel only		Day63	ALH-F1-V fusion	O	O
2086	GP6 Pool	ALH+10µgF1		Day63	ALH-F1-V fusion	40960	O
2087	GP7 Pool	ALH+10µgF2urea		Day63	ALH-F1-V fusion	81920	O
2088	GP8 Pool	ALH+18.5µg F1-V		Day63	ALH-F1-V fusion	81920	327680
2089	GP9 Pool	ALH+37µg F1-V		Day63	ALH-F1-V fusion	163840	163840

AEROSOL EXPOSURE SHEET

[illegible]

u pag 127-128

AEROSOL EXPOSURE SHEET

[illegible]

REDACTED

Exhibit GA5

Dear Dr Friedlander

CBDE/USAMRIID COLLABORATIVE RESEARCH INTO PROTECTIVE EFFICACY OF
RECOMBINANT V-ANTIGEN AGAINST PARENTERAL AND AEROSOL CHALLENGE
WITH YERSINIA PESTIS

As you are aware, CBDE has data to suggest that the V-antigen of the plague causing organism *Yersinia pestis*, when used as an immimogen, is highly protective against plague. The V-antigen could therefore be a major component of an improved plague vaccine to be developed in the future by CBDE.

You recently indicated to us that USAMRIID wished to collaborate in testing the protective capacity of the V-antigen against parenteral and aerosol challenge with virulent plague. We agreed that such a collaboration would be desirable because it could generate valuable data which would be of benefit to both CBDE and USAMRIID. We therefore decided that the collaboration should, in the future, be the subject of a Project Arrangement under the Memorandum of Understanding between the Secretary of Defense (US) and the Secretary of State for Defence (UK) concerning Technology Research and Development Projects (which is currently still under negotiation).

However, we also agreed that any delay in the collaboration would reduce the benefit of the resulting data, and therefore it would be desirable to commence work in advance of a more formal Project Arrangement.

Accordingly, this letter sets out below the respective duties, rights and responsibilities of each of us under the collaboration, *pro tem*, pending the negotiation of a more comprehensive arrangement:

1. SCOPE OF WORK

a. CBDE will supply to USAMRIID, for the purposes described in (b), the following:

- i. 30 mg of recombinant *Yersinia pestis* V-antigen.
- ii. Protocols detailing the immunisation route, doses and schedules used at CBDE.
- iii. Polyclonal antisera raised against the V antigen of *Yersinia pestis*.
- iv. Details of the CBDE challenge route, challenge strain and protection data afforded by the V-antigen vaccine against parenteral challenge with *Yersinia pestis*.

b. USAMRIID will:

- i. Immunise groups of animals parenterally with the following:
 - V-antigen in combination with Alhydrogel.

Suggested protocol for the F1-whole V fusion protein.

File: F1-wholeV fusion last update REDACTED

Protocol: B95-01

F1-WV fusion protein immunization and challenge

Investigators: CPT Heath, Dr. Welkos, LTC Anderson, COL Friedlander

Background. CPT David Heath produced and purified a recombinant F1-V fusion protein. The protein is positive by Western blot to F1 and V. Only part of the V-protein was used in the initial F1-V fusion. This F1-V fusion was immunogenic, but was not very efficacious when compared to the whole V-protein (WV) produced by Mauro to protect against F1 minus strains of *Y. pestis*. This is a repeat of part of the initial F1-V protection study using the whole V-protein in the F1-WV fusion. Subcutaneous injection along back of the mouse for immunization.

Purpose: Immunize and challenge mice to check on immunogenicity and protection against the CO92 and C12 strain of *Y. pestis* by sc and aerosol challenge, **100-Max LD₅₀**.

Alhydrogel, 1.3%, from SuperFos. Batch # 2043, Expiration date None, _____ µg of AL/dose

Endotoxin level in the F1-WV preparation is _____ U/ml.

Will use **Mauro's V** which has been urea treated as per the F1-WV procedure. Details in CPT Heath's laboratory notebook.

Immunization Groups: 10 Swiss Webster female mice per group from Harlan Sprague Dawley

Implantable Micro Identification transponders from: BioMec Data Systems, Inc 255 W. Spring Valley Ave. Maywood, NJ 07607, 1-800-526-BMDS

		Dose		
Subcutaneous challenge		Strain	LD ₅₀	# Mice
Group 1	Alhydrogel alone, days 0, 30, sc	C12	100	10
Group 2	Alhydrogel + 13.6 µg F1-WV fusion protein day 0, 30, sc	C12	100	10 X
Group 3	Alhydrogel + 10 µg Mauro-V urea, days 0, 30, sc	C12	Max	10 X
Group 4	Alhydrogel + 13.6 µg F1-WV fusion protein days 0,30,sc	C12	Max	10 X
Group 5	Alhydrogel + 27.2 µg F1-WV fusion protein days 0, 30, sc	C12	Max	10 X
Group 6	Alhydrogel alone days 0, 30, sc	C12	Max	10 ✓
Group 7	Alhydrogel + 13.6 µg F1-WV fusion protein day 0, 30, sc	CO92	100	10 X
Group 8	Alhydrogel alone, days 0, 30, sc	CO92	100	10

Aerosol challenge

Group 09	Alhydrogel alone, days 0, 30, sc	C12	50	10 ✓
Group 10	Alhydrogel + 13.6 µg F1-WV fusion protein day 0, 30, sc	C12	50	10 X
Group 11	Alhydrogel + 10 µg Mauro-V urea, days 0, 30, sc	C12	Max	10 X
Group 12	Alhydrogel + 13.6 µg F1-WV fusion protein, days 0, 30, sc	C12	Max	10 X
Group 13	Alhydrogel + 27.2 µg F1-WV fusion protein days 0,30, sc	C12	Max	10 X
Group 14	Alhydrogel alone, days 0, 30, sc	C12	Max	10 ✓
Group 15	Alhydrogel + 13.6 µg F1-WV fusion protein days 0, 30, sc	CO92	100	10 X
Group 16	Alhydrogel alone days 0, 30, sc	CO92	100	10 ✓
Group 17	Alhydrogel + 13.6 µg F1-WV fast prep, days 0, 30, sc	C12	Max	10X
Group 18	Alhydrogel + 10 µg F1 + 10 µg Mauro's V, days 0, 30, sc	C12	Max	10 X
Group 19	Greer plague vaccine, days 0, 30, sc	C12	Max	10 05
Group 20	Alhydrogel alone, day 0, 30, sc	C12	Max	05

180

10	Group 21	ALH + 13.6 µg F1-WV fusion protein day 0, 30, sc --antibody response	10
		Measure titer at 7, 14, 27, 57, 90	
	Group 22	ALH + 13.6 µg F1-WV fusion protein day 0, 30, sc --lung lavage, day 57	05
10	Group 23	ALH + 27.2 µg F1-WV fusion protein day 0, 30, sc --antibody response	10
		Measure titer at 7, 14, 27, 57, 90	
	Group 24	ALH + 27.2 µg F1-WV fusion protein day 0, 30, sc --lung lavage, day 57	05
10	Group 25	ALH + Mauro-V urea, 10 ug, day 0, 30, sc --antibody response	10
	Group 26	ALH + Mauro-V urea, 10 ug, day 0, 30, sc --lung lavage, day 57	x 05
	Group 27	ALH alone, day 0, 30, sc	10
		Measure titer at 7, 14, 27, 57, 90	
	Group 28	ALH alone, day 0, 30, sc, lung lavage, day 57	x 05
10	Group 29	ALH alone, for spleen weights 28 day pi	10 05
Total			220

Schedule

Groups 1-20

13Jun95	Arrival of Swiss Webster mice, female 7-8 wks, Harlan Sprague Dawley in AA-3 (Barrier)	
24Jun95	Chipped with BioMedic Data Systems transponders, West	
27Jun95	1st immunization, day 0	
27Jul95	2nd Immunization, day 30	
24Aug95	Bleed to determine prechallenge titers, day 58	
31Aug95	Challenge by aerosol & sc routes, day 65	
28Sep95	Terminal bleed, day 28 pi, titrate spleens#	serum #

Group 21-25

13Jun95	Mice arrive	
27Jun95	1st immunization, AA-3	
11Jul95	Groups 21, 23, 25; Bleed, day 14, AA-3, SERUM#	
26Jul95	Groups 21, 23, 25; Bleed, day 29, AA-3, SERUM#	
27Jul95	2nd immunization, day	
31Aug95	Bleed, day 65, AA-3, Groups	SERUM#
	Groups 22, 24, 26, and 28 for serum titer & bronchial lavage #	
28Sep95	Group 29 for Spleen weights #	
25Sep95	Groups 21, 23, and 25; day 90, serum #	

Chip numbers for all groups extra alhydrogel controls

File: F1-wholeV fusion last update REDACTED
 Protocol: B95-01
 F1-WV fusion protein immunization and challenge

Investigators: CPT Heath, Dr. Welkos, LTC Anderson, COL Friedlander

Background. CPT David Heath produced and purified a recombinant F1-V fusion protein. The protein is positive by Western blot to F1 and V. Only part of the V-protein was used in the initial F1-V fusion. This F1-V fusion was immunogenic, but was not very efficacious when compared to the whole V-protein (WV) produced by Mauro to protect against F1 minus strains of *Y. pestis*. This is a repeat of part of the initial F1-V protection study using the whole V-protein in the F1-WV fusion. Subcutaneous injection along back of the mouse for immunization.

Purpose: Immunize and challenge mice to check on immunogenicity and protection against the CO92 and C12 strain of *Y. pestis* by sc and aerosol challenge, **100-Max LD₅₀**.

Alhydrogel, 1.3%, from SuperFos. Batch # 2043, Expiration date None, 0.1857 mg of AL/dose

Endotoxin level in the F1-WV preparation is _____ U/ml.

Will use Mauro's V which has been urea treated as per the F1-WV procedure. Details in CPT Heath's laboratory notebook.

Immunization Groups: 10 Swiss Webster female mice per group from Harlan Sprague Dawley

Implantable Micro Identification transponders from: BioMeic Data Systems, Inc 255 W. Spring Valley Ave. Maywood, NJ 07607, 1-800-526-BMDS

		Dose		
Subcutaneous challenge		Strain	LD ₅₀	# Mice
Group 1	Alhydrogel alone, days 0, 30, sc	C12	100	10 NC
Group 2	Alhydrogel + 13.6 µg F1-WV fusion protein day 0, 30, sc	C12	100	10
Group 3	Alhydrogel + 10 µg Mauro-V urea, days 0, 30, sc	C12	Max	10
Group 4	Alhydrogel + 13.6 µg F1-WV fusion protein days 0, 30, sc	C12	Max	10
Group 5	Alhydrogel + 27.2 µg F1-WV fusion protein days 0, 30, sc	C12	Max	10
Group 6	Alhydrogel alone days 0, 30, sc	C12	Max	10 NC
Group 7	Alhydrogel + 13.6 µg F1-WV fusion protein day 0, 30, sc	CO92	100	10
Group 8	Alhydrogel alone, days 0, 30, sc	CO92	100	10 NC

Aerosol challenge

Group 09	Alhydrogel alone, days 0, 30, sc	C12	50	10
Group 10	Alhydrogel + 13.6 µg F1-WV fusion protein day 0, 30, sc	C12	50	10
Group 11	Alhydrogel + 10 µg Mauro-V urea, days 0, 30, sc	C12	Max	10
Group 12	Alhydrogel + 13.6 µg F1-WV fusion protein, days 0, 30, sc	C12	Max	10
Group 13	Alhydrogel + 27.2 µg F1-WV fusion protein days 0, 30, sc	C12	Max	10
Group 14	Alhydrogel alone, days 0, 30, sc	C12	Max	10
Group 15	Alhydrogel + 13.6 µg F1-WV fusion protein days 0, 30, sc	CO92	100	10
Group 16	Alhydrogel alone days 0, 30, sc	CO92	100	10
Group 17	Alhydrogel + 10 µg F1 + 10 µg Mauro's V, days 0, 30, sc	C12	Max	10
Group 18	Greer plague vaccine, days 0, 30, sc	C12	Max	09 NC
Group 19	Alhydrogel alone, day 0, 30, sc <i>LIT 1013W2</i>	C12	Max	05

Group 20 ALH + 13.6 µg F1-WV fusion protein day 0, 30, sc --antibody response 10
 Measure titer at X, 14, 27, 57, lung lavage day 57

Pambord/Anderson in AA-4 has 1.8 air exchangers/minute. This is the best when

	Measure titer at X , 14, 27, 57; lung lavage day 57	
Group 22	ALH + Mauro-V urea, 10 ug, day 0, 30, sc --antibody response	10
Group 23	ALH alone, day 0,30, sc	10
	Measure titer at X , 14, 27,57; lung lavage day 57	
Group 24	Greer plague vaccine days 0, 30, sc	
	Measure titer at X , 14, 27, 57; lung lavage day 57 Chipped	05
Group 25	Alhydrogel + 10 µg F1 + 10 µg Mauro's V, days 0, 30, sc	07
	Measure titer at X , 14, 27, 57; lung lavage day 57, Mice rec'd 14Jun95	
	Total	232

Schedule

Groups 1-19

13Jun95 Arrival of Swiss Webster mice, female 7-8 wks, Harlan Sprague Dawley in AA-3 (Barrier)

24Jun95 Chipped with BioMedic Data Systems transponders, West

27Jun95 1st immunization, day 0, Zimmerman, West, Giunanzio, Archer, Anderson

25Jul95 2nd Immunization, day 28, *Hall, West, Zimmerman, Archer, Anderson*

24Aug95 Bleed to determine prechallenge titers, day 58 *5664 - 5847*

08Sep95 Challenge by aerosol & sc routes, day ~~58~~, AA-3 ~~5848-5849~~

28Sep95 Terminal bleed, day 28 pi, titrate spleens# *10030 - 10094* serum # *6082 - 6157*

06 Oct ZIMMERMAN, PLUMMER, ARCHER, SHAMAGIN

Group 20-23

13Jun95 Mice arrive

27Jun95 1st immunization, AA-3

11Jul95 Groups 20, 21, 22; Bleed, day 14, AA-3, SERUM# *4542 - 4593*

26Jul95 Groups 20, 21, 22; Bleed, day 29, AA-3, SERUM# *4807 - 4858 GIUNANZIO,*

25Jul95 2nd immunization, day 28, new barrier, AR-5

04Sep95 Bleed, day 65, AA-3, Groups *24, 25* SERUM# *5848 - 5899 GIUNANZIO, HALL*

 Groups 20, 21, 22, and 23, for serum titer & bronchial lavage #*5900 - 5951*

07Sep95 Group 23 for Spleen weights # *10010 - 10019* *ARCHER, DR PULLMAN*

Chip numbers for all groups extra alhydrogel controls

No Chip GP1

No Chip

No Chip

No Chip

No Chip

No Chip

No Chip

No Chip

No Chip

No Chip

200E2C4363/WV-001 GP2

20103F1F72/WV-002

1F73663C4C/WV-003

1F29341D67/WV-004

2001693A3C/WV-005

2000183C0C/WV-006

1F561F4725/WV-007

1F5F027808/WV-008

1F4B1D3346/WV-009

20023A3371/WV-010

Puncture of peritoneum in AA-4 has 1.8 per exchanges/minute. Take in the box when

No Chip

No Chip

No Chip

No chip

2002537218/WV-051 GP9

201D31583A/WV-052

201D404B38/WV-053

200D22753C/WV-054

1F56211654/WV-055

1F5F154528/WV-056

200A195D60/WV-057

200C043917/WV-058

200142120B/WV-059

1F561F5814/WV-060

201D2D4650/WV-061 GP10

201D30276C/WV-062

1F73085B0B/WV-063

1F757A0A68/WV-064

200F353666/WV-065

203C6C2216/WV-066

200657087B/WV-067

20415E0E33/WV-068

1F501B492D/WV-069

200D7A4D0C/WV-070

1F58195B15/WV-071 GP11

1F7C414D57/WV-072

200A024E06/WV-073

200D34514E/WV-074

1F7F373873/WV-075

2018766969/WV-076

1F61110A65/WV-077

200B0C1732/WV-078

200D51136F/WV-079

1F657E7608/WV-080

20001F4D74/WV-081 GP12

20197C6467/WV-082

1F560B532D/WV-083

2018742331/WV-084

1F28782021/WV-085

203C0C5C3C/WV-086

1F402E787B/WV-087

200B386439/WV-088

200D7E7D58/WV-089

200666175D/WV-090

2019711D39/WV-091 GP12

20017C5211/WV-092

201B493C40/WV-093

1F4E1D383E/WV-094

200B691F4D/WV-095

1F63265C7C/WV-096

1F56147502/WV-097

2041632517/WV-098

Weight & time of challenge

34.2

29.0

34.4

41.9

29.8

37.5

28.5

28.3

34.9

37.4

$335.9 / 10 = 33.6 \text{ gm avg. weight}$

Pumping/handflow in AA-4 has 1.8 per syphonage/minute. Then in the box where

1F720E412D/WV-101 GP14
1F8A100031/WV-102
2007407200/WV-103
200F44711D/WV-104
20440E5A84/WV-105
201D452F4F/WV-106
2000093A20/WV-107
1F707C4227/WV-108
2035422049/WV-109
201D2E2C69/WV-110
203A555100/WV-111 GP15
1F561E2E3F/WV-112
1F57015930/WV-113
1F5626184D/WV-114
200F31011F/WV-115
201E6C4313/WV-116
2018704C0C/WV-117
1F19690758/WV-118
200F136D51/WV-119
1F76132731/WV-120
201052314D/WV-131 GP16
1F5615492D/WV-132
1F73656128/WV-133
2001311618/WV-134
20104C6E16/WV-135
201C317E15/WV-136
200D48117A/WV-137
200046504A/WV-138
1F62746427/WV-139
201D483546/WV-140
201D2E2A6B/WV-141 GP17
200A05272A/WV-142
2000137756/WV-143
200C06410D/WV-144 - 1F6C7F3343
1F7D60453F/WV-145
2041580443/WV-146
1F75787C78/WV-147
1F20264457/WV-148
1F560D116D/WV-149
1F73671E69/WV-150
No chip GP18
No chip
No chip
No chip
No chip
No chip
No chip
No chip
No chip
2000037B62/WV-122 GP19
1F6679384A/WV-123
20093B4D4F/WV-125

On low level building in AA-4 has 1.8 per up/down / minute. Then in the log when

1F5010000/WV-155 GP20
 1F5010000/WV-157
 1F5010000/WV-158
 1F5010000/WV-159
 1F5010000/WV-160
 1F5010000/WV-161
 1F5010000/WV-162
 1F5010000/WV-163
 1F5010000/WV-164
 1F5010000/WV-165
 1F5010000/WV-166 GP21
 1F5010000/WV-167
 200A112F16/WV-168
 1F55244324/WV-169
 200D2F170D/WV-170
 20097A0657/WV-161
 200C032829/WV-162
 2010346339/WV-163
 1F743C6948/WV-164
 201F012818/WV-165
 201D480C8F/WV-176 GP22
 2035323B3E/WV-177
 1F29476B06/WV-178
 1F791B202D/WV-179
 20415D784A/WV-180
 203C137F12/WV-171
 1F60390543/WV-172
 1F76001259/WV-173
 1F56035E2A/WV-174
 20447C326E/WV-175
 1F7C4C0217/WV-186 GP23
 1F203F156D/WV-187
 2001361415/WV-188
 1F7C2B0733/WV-189
 1F73571B7C/WV-190
 1F6337794E/WV-181
 200B5E4C2B/WV-182
 1F60786524/WV-183
 20024F226D/WV-184
 1F56155B1B/WV-185
 200F6B1551/WV-121 GP24
 1F2052056A/WV-124
 2009221C19/WV-126
 1F2A6A1E2F/WV-128
 2008425640/WV-130
 No Chip GP25
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pin 20429 95, came unknown

Pumped/flowline in AA-4 has 1.8 psi up/down/mo. Taken in the box when

Project: Recombinant F1-wholeV Fusion	
Notebook #: 3739	
Inoculum: Yersinia pestis strain, CO92	
Route: sc	Dose:
Swiss Webster	Arri
7-8wks	Vendor: Harlan Sprague Dawley
Sex: female	
Day postinfection	0 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29
Group	LD50
1 C12	Alhydrogel alone
2 C12	Alhydrogel 13.6ug F1-WV
3 C12	Alhydrogel 10ug Mauro-V
4 C12	Alhydrogel 13.6ug F1-WV
5 C12	Alhydrogel 27.2ug F1-WV
6 C12	Alhydrogel alone
For Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions Discard dead animals Use scanner to check chip number of dead mice Mark number of animals alive in each cage re = mouse has been rechipped	

REDACTED

Project: Recombinant F1-wholeV Fusion

tebook #: 3739

cculum: Yersinia pestis strain, CO92 OR C12

ute: AEROSOL

Dose:

EDACTED

Wiss Webster

REDACTED

at 7-8wks

Vendor: Harlan Sprague Dawley

Sex: female

ay -

y postinfection

oup

LD50

9 C12

hydrogel

me

BLACK

WHITE

WISSD 303a0

10 C12

hydrogel

.6ug

-WV

WHITE

WHITE

RUN 1

11 C12

hydrogel

Max

ug

uro-v

ia

WHITE

WISSD 6.0114

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WISSD 6.0114

WISSD 6.0114

Comments/Chip #

2002337219/WV-051

201D31583A/WV-052

201D404B98/WV-053

200D22753C/WV-054

1F50211054/WV-055

1F50F154528/WV-056

200A195D66/WV-057

200G040917/WV-058

2001424208A/WV-059

1F561F5014/WV-060

201D2D4650/WV-061

201D30276C/WV-062

1F73085B0B/WV-063

1F757A0A68/WV-064

200F353666/WV-065

203C6C2216/WV-066

200657087B/WV-067

20415E0E33/WV-068

1F501B492D/WV-069

200D7A4D0C/WV-070

1F50195B15A/WV-071

1F7C414D57/WV-072

200A024E06/WV-073

200D34514E/WV-074

1F7F373873/WV-075

2018766969/WV-076

1F61110A65/WV-077

200B0C1732/WV-078

200D51136F/WV-079

1F657E7608/WV-080

20001F4D74/WV-081

20197C6467/WV-082

1F560B532D/WV-083

2018742331/WV-084

1F28782021/WV-085

203C0C5C3C/WV-086

1F402E787B/WV-087

200B386439/WV-088

200D7E7D58/WV-089

200666175D/WV-090

2019711D39/WV-091

20017C5211/WV-092

201B493C40/WV-093

1F4E1D383E/WV-094

200B691F4D/WV-095

1F63265C7C/WV-096

1F56147502/WV-097

2041832517/WV-098

1F560E502D/WV-099

1F7D1A5E6C/WV-100

1F720E4120/WV-101

1F6A163031A/WV-102

200749726E/WV-103

200F44711C/WV-104

20440E5A54/WV-105

201D452F4F/WV-106

2003033A20/WV-107

1F7C7C4227/WV-108

2005422049/WV-109

201D2E2C09/WV-110

Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions

card dead animals

3 scanner to check chip number of dead mice

rk number of animals alive in each cage

= mouse has been rechipped

File: F1-V, longterm Last update: REDACTED
 Protocol: B95-01
 Investigators: Anderson/Heath/Welkos/Friedlander

Background: F1-WV and F1 and V in combination have been shown to protect against challenges of CO92 and C12 with a two dose schedule (0 and 30). The long-term decay of the antibody response to the initial immunization and length of protection from a single immunization is currently not known.

Purpose: To examine the decay of the antibody response to an initial immunization, protection afforded over time to an initial immunization to indicate the optimum time for the 2nd immunization for an aerosol challenge. Titers to F1 and V will be determined. Mice will be challenged with 50-100 LD₅₀ CO92, aerosol challenge.

In the below challenge groups, when protection falls to zero, the remaining groups will be booster and challenged 2 weeks post-boost.

Immunogens: EcF1s, Mauro's V, and F1-WV all essentially endotoxin free from Dr. Heath.

Mice: Swiss Webster (Hsd:ND4) female mice per group from Harlan Sprague Dawley, Inc. Indianapolis, Indiana (317) 894-7521.

Alhydrogel: 1.3%, from SuperFos. Batch # 2043, Expiration date None, 0.1857 mg of AL/dose

IMI-1000, Implantable micro identification transponders, BioMedic Data Systems, Inc., 255 W. Spring Valley Ave. Maywood, NJ 07607, Tel 201 587-8300.

Plague USP, Lot # 112FX1, Expiration Date REDACTED, Greer Laboratories, Inc. P.O. Box 800 Lenoir, NC 28645-0800

Group	Treatment	Challenge Day	# of mice	
1A	10µF1+20µgMauro-V day0	14	10	<i>serum # 6990-7039 - pre challenge</i> <i>serum # 10146-10160 see page 81, 85</i> <i>serum # 7820-7833 day 28 PI post challenge</i>
1B			05	
2A	30µgF1-WV	14	10	
2B			05	
3	Plague USP	14	10	
4	Alhydrogel alone	14	10	
5A	10µF1+20µgMauro-V day0	42	10	<i>serum # 7676-7725 pre challenge</i> <i>serum # 10160-10191 day 30 PI</i> <i>serum # 8004-8035 day 30 PI post challenge</i>
5B			05	
6A	30µgF1-WV	42	10	
6B			05	
7	Plague USP	42	10	
8	Alhydrogel alone	42	10	
9A	10µF1+20µgMauro-V day0	<u>98 119</u>	10	<i>serum # 8288-8337 pre challenge</i> <i>serum # 8591-8621 day 28 PI</i> <i>serum # 10259-10290 day 28 PI</i>
9B			05	
10A	30µgF1-WV	<u>98 119</u>	10	
10B			05	
11	Plague USP	<u>98 119</u>	10	
12	Alhydrogel alone	<u>98 119</u>	10	

13A 10μF1+20μgMauro-V day0
 13B
 14A 30μgF1-WV
 14B
 15 Plague USP
 16 Alhydrogel alone

10
 05
 10
 05
 10
 10

DMT # 0416-2753

17A 10μF1+20μgMauro-V day0
 17B
 18A 30μgF1-WV
 18B
 19 Plague USP
 20 Alhydrogel alone

10 serial bleeds on challenge days,
 05
 10 14, 42, 93, and
 05 day 14 serum # 7040-5097
 10 42 # 7822-7823 774-7819
 10 93 # 8400-8449

challenge
 day 358
 730696

Total 250

204 # 8906-8955
 250 # 8545-8577 180696

Schedule

17Oct95 Mice arrive B412
 25Oct95 Mice chipped B412, Plumtree, Zimmerman
 31Oct95 Mice immunized day 0, Plumtree, Zimmerman
 07Nov95 Bleed 1st challenge group, day 7 serum #
 14Nov95 1st challenge group, day 14, 100 LD₅₀ aerosol challenge, CO92
 14Nov95 Bleed serial bleed group, day 14
 05Dec95 Bleed 2nd challenge group, day 35
 12Dec95 2nd challenge group, day 42, 100 LD₅₀ aerosol challenge, CO92
 12Dec95 Bleed serial bleed group, day 42
 25Jan96 Bleed 3rd challenge group, 86 DAY PLUMTREE, SAMBAIN, IMMUNIZED
 01Feb96 3rd challenge group, day 93, 100 LD₅₀ aerosol challenge, CO92
 01Feb96 Bleed serial bleed group, day 93
 _____95 Bleed 4th challenge group, day
 _____96 4th challenge group, day, 100 LD₅₀ aerosol challenge, CO92
 _____96 Bleed serial bleed group, day

Chips Numbers

22254D6722/LT-001 GP1A
 22254B4164/LT-002
 221D487E7E/LT-003
 221D705617/LT-004
 22213D0B42/LT-005
 222122186F/LT-006
 22223E1239/LT-007
 2221493775/LT-008
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 22213D5220/LT-010
 222233166C/LT-011 GP2A
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 221D6C477D/LT-013
 2222485A77/LT-014
 221A2B093D/LT-015
 221D6B7917/LT-016
 221D686E17/LT-017
 2222411328/LT-018
 22225C6018/LT-019
 221D630D5E/LT-020
 221D4E2460/LT-021 GP3

22280F622F/LT-022
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221D552A40/LT-024
22223C1767/LT-025
2221534945/LT-026
22277D234D/LT-027
221D660734/LT-028
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222243493A/LT-075

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22224C4154/LT-126
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22223A6E6D/LT-197
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22215A7309/LT-205
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221D49253C/LT-207
221D710C76/LT-208
22273F017C/LT-209
22217E5E6E/LT-210
221D494F34/LT-211 GP5B
22217B022D/LT-212
2221386837/LT-213
2228057813/LT-214
222142434F/LT-215
2222323813/LT-216 GP6B
222121405D/LT-217
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2225527970/LT-219
221D56403B/LT-220
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22280E312B/LT-222
221D584D78/LT-223
221D4C4067/LT-224
22212A2966/LT-225
222124374C/LT-226 GP10B
2222586303/LT-227
2222371808/LT-228
2222443D5B/LT-229
22213B707C/LT-230
221D737744/LT-231 GP13B
2221313035/LT-232
2222392702/LT-233
221D73716A/LT-234
221D527D35/LT-235
221B2E7B71/LT-236 GP14B
221D484645/LT-237

22740000/LT-238
2210400000/LT-239
2202010000/LT-240
2221000000/LT-241 GP17B
2210000040/LT-242
2221400071/LT-243
2210000000/LT-244
2222400070/LT-245
1F7E000077/LT-246 GP18B
2222400004/LT-247 - 1.07 AMP, RESISTANCE 1F7E1B695F
2221470734/LT-248
22254A3C0B/LT-249
221A422E0B/LT-250
1F72777B7D/TESTCHIP

Exhibit GA11

File: Alhydrogel concentration

Last updated: REDACTED

Anderson/Heath/Welkos/Friedlander

Background. The allowable Al content in a human vaccine is 0.85 mg/dose as determined by assay (21 CFR 610.15(1)). However, the lowest possible dose of Al should be used which maintains an adequate adjuvant response with the EcF1c and V immunogens. A dose response for Alhydrogel has not been done with a combination of F1 and V. Therefore this experiment will examine a range of concentrations of AL which will be used with a constant amount of EcF1c and V. EcF1c (60 EU/ml) and V-His tag (preparation are essentially endotoxin free. The level of endotoxin in the thrombin treated V preparation without the His tag is 49 EU/ml.

Compare the antibody response to V with F1-WV protein with and without alhydrogel. When F1 + V is used to immunize without alhydrogel, there was not antibody response to V. This will be done with F1-WV in order to determine whether F1 is contributing anything to protection with the F1-WV protein. F1-WV purified by Ni++ and ran through a Sartorius Q15 filter using 10mM Tris, pH 7.6, 0.5mM EDTA + 0.5 MNaCl for elution. See Heath's note of 5-6 Dec. Endotoxin ____ EU/ml.

Mice: Swiss Webster (Hsd:ND4) female mice from Harlan Sprague Dawley, Inc. Indianapolis, Indiana (317) 894-7521.

1.3% Alhydrogel (Aluminium Hydroxide Gel Adjuvant): Al_2O_3 (1.3%) equivalent to $Al(OH)_3$ (2.0%), from SuperFos Biosector a/s, Frydeniundsvej 30, DK-2950 Vedbaek, Denmark. Batch # 2043, Expiration date None; U.S. supplier - Accurate Chemical & Scientific Corp, 300 Shames Drive, Westbury, NY 11590, Tel (516) 333-2221, Fax (516) 997-4948.

Al = 13 O = 8 H = 1, $Al(OH)_3$ = 40 molecular weight

Current procedure for adsorption of F1 and V to Alhydrogel: 1.0ml Alhydrogel brought to 7.0 ml
2% $Al(OH)_3$ = 20 mg/ml

$20mg/ml(1.0ml) = (x)(7.0ml \text{ final volume})$ $x = 2.857 \text{ mg } Al(OH)_3/ml$

$(2.857mg/ml)(0.2ml \text{ dose}) = 0.57142 \text{ mg } Al(OH)_3$

AL is 0.325% of $Al(OH)_3$

0.235% of 2.857mg = 0.1857 mg of AL/0.2ml dose which the mouse receives

Current dose of Al which has been used through out the mouse experiments is 0.1857mg. Try two other doses each 75% of the former GP1 = 0.1857mg (standard amount of AL), GP2 = 0.1393mg, GP3 = 0.1045mg

IMI-1000, Implantable micro identification transponders, BioMedic Data Systems, Inc., 255 W. Spring Valley Ave. Maywood, NJ 07607, Tel 201 587-8300.

Plague USP, Lot # 1128X1, Expiration Date REDACTED, Greer Laboratories, Inc. P.O Box 800 Lenoir, NC 28645-0800,

Groups	Treatment	V-HIS Tag	Strain	#Mice
000A	30µgF1-WV, no AL	Yes	CO92	10
000B	30µgF1-WV, no AL	Yes	CO92	05
00A	30µgF1-WV+0.19Al	Yes	CO92	10
00B	30µgF1-WV+0.19Al	Yes	CO92	05
0A	10µgF1+20µgV+0.19Al	No	CO92	10
0B	10µgF1+20µgV+0.19Al	No	CO92	10

Total 210

serum # spleen #

2225535315/ALH-032

See pages 104-107

2222382716/ALH-033
221D64205D/ALH-034
2221361744/ALH-035
221B3B756D/ALH-041 GP0A
2222505C62/ALH-042
221D355F0B/ALH-043
2221393824/ALH-044
2221260562/ALH-045
2221750728/ALH-046
2221710C75/ALH-047
22254A533A/ALH-048
221D48425F/ALH-049
222547047F/ALH-050
221B431E0B/ALH-051 GP0B
222141267F/ALH-052
2225447977/ALH-053
22217C5904/ALH-054
22276E4F74/ALH-055
2222513264/ALH-056
221B331A0B/ALH-057
22223A2C20/ALH-058
2225301E16/ALH-059
2222464F0C/ALH-060
2225525E7E/ALH-061 GP1A
22214B6521/ALH-062
2225427F32/ALH-063
221B2E1A28/ALH-064
22277F2A4F/ALH-065
221B381564/ALH-066
22276C4058/ALH-067
221B365521/ALH-068
2221483A1E/ALH-069
221B2F4049/ALH-070
2222490E7E/ALH-071 GP1B
22216B6702/ALH-072
22281D7640/ALH-073
221D74327B/ALH-074
221B36013C/ALH-075
2227177522/ALH-076
221B560D16/ALH-077
222734024F/ALH-078
221B463462/ALH-079
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22252D6D26/ALH-081 GP2A
2225416D0B/ALH-082
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222550400F/ALH-085
221B496E3F/ALH-086
2221353A4B/ALH-087
221D542860/ALH-088
2225376005/ALH-089
221B3A5805/ALH-090
221D74480D/ALH-091 GP2B

from USAMRIID notebook

*From USMC 100 military
of Lane North*

2225315130/ALH-093
2221630F69/ALH-094
221D615D3E/ALH-095
2225574367/ALH-096
221B440F29/ALH-097
222176302B/ALH-098
22212E212C/ALH-099
22273B1603/ALH-100
22274C0115/ALH-101 GP3A
222178537A/ALH-102
2225447B43/ALH-103
2221775C19/ALH-104
2227351528/ALH-105
22271B745A/ALH-106
2221525A38/ALH-107
2225546B51/ALH-108
2225316828/ALH-109
222243260B/ALH-110
2221694D14/ALH-111 GP3B
22277D2440/ALH-112
2225407A6C/ALH-113
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2227514F27/ALH-116
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22254F0206/ALH-118
221B3B1F65/ALH-119
222543090C/ALH-120
2225402D66/ALH-121 GP4
2228015179/ALH-122
2225343914/ALH-123
222529444C/ALH-124
22271B7357/ALH-125
222732682C/ALH-126
2221501931/ALH-127
22196E7321/ALH-128
22280F5F2D/ALH-129
222551747C/ALH-130
2222464E2F/ALH-131 GP5
22272E117A/ALH-132
22225C6636/ALH-133
22254E3756/ALH-134
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2225572C56/ALH-138
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221D6A064C/ALH-141 GP6
22254C574B/ALH-142
22224D0523/ALH-143
2225561B32/ALH-144
2227475E7D/ALH-145

See pages 104-107

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2221702733/ALH-146
221D504A22/ALH-147
221B330124/ALH-148
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22253B326D/ALH-150
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2221641439/ALH-195
2225443B3A/ALH-196
2222585764/ALH-197
22213B7E14/ALH-198
222160194A/ALH-199

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the pages 104-107

2221490005/ALH-200
22213D1B3C/ALH-201 GP12
2222523865/ALH-202
2221707A4B/ALH-203
221B41083C/ALH-204
222837504A/ALH-205
22273A4C62/ALH-206
22216B1128/ALH-207
221B511B39/ALH-208
2222557D5A/ALH-209
221B432713/ALH-210 GP13
221D73342A/ALH-016
221D492B54/ALH-017
2228145E46/ALH-018
221D763028/ALH-019
2221543C07/ALH-020
22225A6150/ALH-036
2221534A52/ALH-037
221D73450D/ALH-038
22276D5C6E/ALH-039
2227557710/ALH-040

see page 104-107

REDACTED

From USMA110 notebook
of Dave Heath

2221701/44D/ALH-205
221B41083C/ALH-204
222837504A/ALH-205
22273A4C62/ALH-206
9916R119R/ALH-207

DACTED

SET UP ABSORPTIONS FOR LTC ANDERSON:

GRPS 000A + 000B used FI-V that had been purified by Ni^{2+} & run through a Sartorius Q15 filter using 10 mM Tris, pH 7.6 0.5 mM EDTA + 0.5 M NaCl for elution. Buffer exchanged this using 10 mM Tris, pH 7.6 + 0.5 mM EDTA using a centrifuge. Did BCA assay for protein conc. = 690 $\mu\text{g}/\text{mL}$ and 1.5 μL total volume

so used 450 μg in 3 mL PBS = 652 μL of FI-V (@ 690 $\mu\text{g}/\text{mL}$)
652 μL FI-V + 2.348 mL PBS

GRPS 00A same FI-V as above @ 690 $\mu\text{g}/\text{mL}$
00B add 652 μL of FI-V to 428 mL ALKHYDROGEL + ABSORBED FOR 2 HR @ 4°C

- spun tube @ 2000 rpm for 5 min. & removed 2 100 μL aliquots to check for protein conc. on BCA assay (see BCA assay results)

- added PBS to absorbed alk-FI-V to 3 mL

PS 0A + 0B Took FI capsule extract @ 60 $\mu\text{g}/\text{mL}$ & conc. of 705 $\mu\text{g}/\text{mL}$ & added 496 μL of this to 1 mL ALKHYDROGEL + 137 μL of threonine treated Vmaxa (5.1 $\mu\text{g}/\text{mL}$) = 496 $\mu\text{L}/\text{mL}$
this is for 10 $\mu\text{g}/100 \mu\text{L}$ dose of FI + 20 $\mu\text{g}/100 \mu\text{L}$ dose V in a final volume of 7 mL

496 μL of FI = 350 μg 137 μL V = 700 μg V

496 μL FI + 137 μL threonine V + 267 μL PBS + 1 mL ALKHYDROGEL

REDACTED

(cont)

GRPS 1A + 1B 10mg FI + 20mg hV in 7ml final volume $hV = 1.36U/ml$
496ul FI (705mg/ml) + 129ul hV (514mg/ml) + 375ul PBS
+ 1ml ALHYDROGEN, Rock BV @ 40C

- Spin 2000 rpm, 5 min. remove 2 x 100ul aliquots for BCA Assay of absorption
- Then added PBS to 7ml final volume

GRPS 2A + 2B Same as 1A + 1B above but used 750ul ALHYDROGEN
+ additional 250ul PBS

GRPS 3A + 3B Same as 1A + 1B above but used 562.5ul ALHYDROGEN
+ 437.5ul PBS to equal 1ml then same as 1A + 1B

GRPS 4 + 5 Added 1ml ALHYDROGEN to 1ml PBS + Rocked BV. Then
added PBS to 7ml final volume

GRPS 6 added 750ul ALHYDROGEN + 250ul PBS + 1ml PBS +
rocked BV @ 40C. Then added PBS to 7ml
final volume.

GRPS 7 added 562.5ul ALHYDROGEN + 437.5ul PBS then 1ml PBS
+ Rocked BV @ 40C
- added PBS to 7ml final volume

GRPS 8 + 9 Added 496ul FI + 129ul hV + 1.375ml PBS + Rocked BV.
Then added PBS to 7ml final volume

0011

LOST after Chip # means the chip has fallen out

FEV_P - MUST COME OUT OF INITIAL PERIOD OF CALLING

Project: F1+V alhydrogel concentrations, V w&wo His tag, F1-WV w&wo alhydrogel	
Notebook #: 3739	
Medium: <i>Yersinia pestis</i> strain, CO92	
Route: aerosol Dose: as shown below <i>LD50 191-304</i>	
Swiss Webster	at 7-8wks
Vendor: Harlan Sprague Dawley	Sex: female
REDACTED	REDACTED
Day -	19 20 21 22 23 24 25 26 27 28 29
Day postinfection	0 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29
Group	LD50
Plague (Greer) s.d.	CO92
1128X1	<i>DID NOT CHALLENGE</i>
11	CO92
Plague (Greer) s.d.	CO92
1128X1	<i>DID NOT CHALLENGE</i>
12	CO92
No Treatment	
13	CO92
Plague (Greer) l.m.	CO92
1128X1	<i>DID NOT CHALLENGE</i>
CO92	
CO92	

For Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions

Discard dead animals

Check chip number of dead mice with scanner

Mark number of animals alive in each cage

LOST after Chip # means the chip has fallen out

low score 88

AEROSOL EXPOSURE SHEET

Aerosol exposure #: 56-016 H	Location: AP-4 H	Date: 23 Feb 96
Aerosol Operator: JPK Adams		Agent: Plague CO92
Pre-operational Check Performed: <input checked="" type="checkbox"/>	Dry T: 71 Wet T: 63 Rel. Hum.: 63%	Protocol #:
Exposure System Type/ID: New Only		P.I.: COL Byrne
System Flow Rate: 12.0 LPM	Timer Check: <input checked="" type="checkbox"/> Pre <input checked="" type="checkbox"/> Post	LTC Anderson
Collision #: A	Start time 0930 1400	
Panel #: 3	End time 0935 1407	
Electronic Flow Meter #: R-0979		

Run #	Animal #	Animal Species	Start T			5 min T			Start Time	AGI #	Comments
			Dry	Wet	Rel. Hum.	Dry	Wet	Rel. Hum.			
1	25	Mice	71	63	63%	71	67	80%	1001	52	
2	25	Mice	73	63	58%	73	68	78.5%	1030	1163	* Cannot Lower Humidity, because
3	25	Mice	73	63	58%	73	68	78.5%	1054	361	all air bubbles to secondary air
4	25	Mice	73	63	58%	73	68	78.5%	1119	26	air turned off. (Bust)
5	23	Mice	74	63	55%	74	68	68%	1152	360	
6	25	Mice	75	64	57%	75	68	70%	1222	72	
7	25	Mice	75	63	51%	74	66	68%	1321	5	* One mouse dead on arrival
											B- J. H. 23 Feb 96

Antibiotic Treatment / Vaccine Challenge

Date: 23-Feb

PI: COL Byrne / LTC Anderson
Agent: Plague
Strain: CO92

Animal Model: Mouse

Strain: Swiss Webster

Wt: (Ave.) : 20g / 29.98g

CO92 LD50=2.1E+04

↓
for runs 6 & 7, Anderson

Sex: female

Run #	AGI/ml	AGI	cfu/l aerosol	MV	Inhaled Dose cfu	LD50s	
1	3.50E+07	3.50E+08	5.83E+06	0.02	1.17E+06	55.56	CO92
2	3.80E+07	3.80E+08	6.33E+06	0.02	1.27E+06	60.32	CO92
3	5.10E+07	5.10E+08	8.50E+06	0.02	1.70E+06	80.95	CO92
4	4.00E+07	4.00E+08	6.67E+06	0.02	1.33E+06	63.49	CO92
5	8.10E+06	8.10E+07	1.35E+06	0.02	2.70E+05	12.86	CO92
6	1.70E+07	1.70E+08	2.83E+06	0.027	7.65E+05	36.43	CO92
7	1.60E+07	1.60E+08	2.67E+06	0.027	7.20E+05	34.29	CO92

[illegible]

ELISA V Summary *Le page 75 4102* 3 APR 96

PROTOCOL: V LONGTERM				V antigen TITER
Plate	Serum	Group Treatment	Bleed	IgG(H+L)
1A	8288	GP9A F1+V	+DAY86	81,920
1B	8289	GP9A F1+V	+DAY86	327,680
2A	8290	GP9A F1+V	+DAY86	40,960
2B	8291	GP9A F1+V	+DAY86	NO SERUM
3A	8292	GP9A F1+V	+DAY86	163,840
3B	8293	GP9A F1+V	+DAY86	327,680
4A	8294	GP9A F1+V	+DAY86	327,680
4B	8295	GP9A F1+V	+DAY86	655,360
5A	8296	GP9A F1+V	+DAY86	40,960
5B	8297	GP9A F1+V	+DAY86	655,360
6A	8298	GP9B F1+V	+DAY86	327,680
6B	8299	GP9B F1+V	+DAY86	327,680
7A	8300	GP9B F1+V	+DAY86	81,920
7B	8301	GP9B F1+V	+DAY86	655,360
8A	8302	GP9B F1+V	+DAY86	327,680
Geomean				220,512

8B	8303	GP10A F1-WV	+DAY86	163,840
9A	8304	GP10A F1-WV	+DAY86	163,840
9B	8305	GP10A F1-WV	+DAY86	163,840
10A	8306	GP10A F1-WV	+DAY86	163,840
10B	8307	GP10A F1-WV	+DAY86	163,840
11A	8308	GP10A F1-WV	+DAY86	81,920
11B	8309	GP10A F1-WV	+DAY86	163,840
12A	8310	GP10A F1-WV	+DAY86	163,840
12B	8311	GP10A F1-WV	+DAY86	163,840
13A	8312	GP10A F1-WV	+DAY86	163,840
13B	8313	GP10B F1-WV	+DAY86	81,920
14A	8314	GP10B F1-WV	+DAY86	327,680
14B	8315	GP10B F1-WV	+DAY86	327,680
15A	8316	GP10B F1-WV	+DAY86	163,840
15B	8317	GP10B F1-WV	+DAY86	327,680
Geomean				171,589

16A	8318	GP11 PLAGUE USP	+DAY86	1,280
16B	8319	GP11 PLAGUE USP	+DAY86	640
17A	8320	GP11 PLAGUE USP	+DAY86	1,280
17B	8321	GP11 PLAGUE USP	+DAY86	640
18A	8322	GP11 PLAGUE USP	+DAY86	1,280
18B	8323	GP11 PLAGUE USP	+DAY86	1,280
19A	8324	GP11 PLAGUE USP	+DAY86	1,280
19B	8325	GP11 PLAGUE USP	+DAY86	640
20A	8326	GP11 PLAGUE USP	+DAY86	540
20B	8327	GP11 PLAGUE USP	+DAY86	1,280
Geomean				970

21A	8328	GP12 ALHYDRO ALONE	+DAY86	640
21B	8329	GP12 ALHYDRO ALONE	+DAY86	640
22A	8331	GP12 ALHYDRO ALONE	+DAY86	640
22B	8333	GP12 ALHYDRO ALONE	+DAY86	1,280
23A	8335	GP12 ALHYDRO ALONE	+DAY86	1,280
23B	8336	GP12 ALHYDRO ALONE	+DAY86	1,280
24A	8337	GP12 ALHYDRO ALONE	+DAY86	640
Geomean				861

24B F1/V POOL

655,360

ELISA F1 Summary

5 APR 96

PROTOCOL: V/F1 LONGTERM				F1 antigen TITER
Plate	Serum	Group Treatment	Bleed	IgG(H+L)
1A	8288	GP9A F1+V	+DAY86	NSUFFICIENT SERUM
1B	8289	GP9A F1+V	+DAY86	20,480
2A	8290	GP9A F1+V	+DAY86	NSUFFICIENT SERUM
	8291	GP9A F1+V	+DAY86	NO SERUM
2B	8292	GP9A F1+V	+DAY86	40,960
3A	8293	GP9A F1+V	+DAY86	2,560
3B	8294	GP9A F1+V	+DAY86	20,480
4A	8295	GP9A F1+V	+DAY86	5,120
4B	8296	GP9A F1+V	+DAY86	20,480
5A	8297	GP9A F1+V	+DAY86	81,920
5B	8298	GP9B F1+V	+DAY86	40,960
6A	8299	GP9B F1+V	+DAY86	2,560
6B	8300	GP9B F1+V	+DAY86	20,480
7A	8301	GP9B F1+V	+DAY86	40,960
7B	8302	GP9B F1+V	+DAY86	5,120
Geomean				15,343

8A	8303	GP10A F1-WV	+DAY86	5,120
8B	8304	GP10A F1-WV	+DAY86	20,480
9A	8305	GP10A F1-WV	+DAY86	1,280
9B	8306	GP10A F1-WV	+DAY86	10,240
10A	8307	GP10A F1-WV	+DAY86	10,240
10B	8308	GP10A F1-WV	+DAY86	2,560
11A	8309	GP10A F1-WV	+DAY86	2,560
11B	8310	GP10A F1-WV	+DAY86	5,120
12A	8311	GP10A F1-WV	+DAY86	1,280
12B	8312	GP10A F1-WV	+DAY86	5,120
13A	8313	GP10B F1-WV	+DAY86	1,280
13B	8314	GP10B F1-WV	+DAY86	2,560
14A	8315	GP10B F1-WV	+DAY86	5,120
14B	8316	GP10B F1-WV	+DAY86	5,120
15A	8317	GP10B F1-WV	+DAY86	10,240
Geomean				4,256

15B	8318	GP11 PLAGUE USP	+DAY86	5,120
16A	8319	GP11 PLAGUE USP	+DAY86	5,120
16B	8320	GP11 PLAGUE USP	+DAY86	10,240
17A	8321	GP11 PLAGUE USP	+DAY86	10,240
17B	8322	GP11 PLAGUE USP	+DAY86	10,240
18A	8323	GP11 PLAGUE USP	+DAY86	5,120
18B	8324	GP11 PLAGUE USP	+DAY86	10,240
19A	8325	GP11 PLAGUE USP	+DAY86	20,480
19B	8326	GP11 PLAGUE USP	+DAY86	5,120
20A	8327	GP11 PLAGUE USP	+DAY86	10,240
Geomean				8,317

20B	8328	GP12 ALHYDRO ALONE	+DAY86	320
21A	8329	GP12 ALHYDRO ALONE	+DAY86	320
21B	8331	GP12 ALHYDRO ALONE	+DAY86	320
22A	8333	GP12 ALHYDRO ALONE	+DAY86	320
22B	8335	GP12 ALHYDRO ALONE	+DAY86	320
23A	8336	GP12 ALHYDRO ALONE	+DAY86	320
23B	8337	GP12 ALHYDRO ALONE	+DAY86	320
Geomean				320

24A F1/V POOL

40,960

500ml, undiluted

9 APR 96

Hbt	Hbt	Hbt	Hbt	Hbt	Hbt	Hbt	Hbt	Hbt	Hbt
No: 1	No: 2	No: 3	No: 4	No: 5	No: 6	No: 7	No: 8	No: 9	No: 10
1	1	1	1	1	1	1	1	1	1
2 ^a	2 ^a	2 ^a	2 ^a	2 ^a	2 ^a	2 ^a	2 ^a	2 ^a	2 ^a
3	3	3	3	3	3	3	3	3	3
A	A	A	A	A	A	A	A	A	A
E	E	E	E	E	E	E	E	E	E
M	M	M	M	M	M	M	M	M	M
K	K	K	K	K	K	K	K	K	K
λ	λ	λ	λ	λ	λ	λ	λ	λ	λ

On serum samples
all 2640 mm on
2 may 96

MAB V-YP 7F5-1-1, SUBCLONES -10

Mean time-to-death (MTD)																
Challenge group	1	2	3	4	5	6	7	8	9	10	MTD	+/- Stdev	Std error	Group		
Subcutaneous:																
1 alhydrogel alone, days 0, 30, sc																
C12 100	5	6	6	7	7	8	8	8	10	11	7.6	1.837873	0.61262	1		
2 alhydrogel + 13.6 µg F1-WV fusion protein day 0, 30, sc																
C12 100	28	28	28	28	28	28	28	28	28	28	28	0	0	2		
3 alhydrogel + 10 µg Mauro-V urea, days 0, 30, sc																
C12 Max	6	28	28	28	28	28	28	28	28	28	25.8	6.957011	2.319	3		
4 alhydrogel + 13.6 µg F1-WV fusion protein day 0, 30, sc																
C12 Max	6	28	28	28	28	28	28	28	28	28	25.8	6.957011	2.319	4		
5 alhydrogel + 27.2 µg F1-WV fusion protein day 0, 30, sc																
C12 Max	28	28	28	28	28	28	28	28	28	28	28	0	0	5		
6 alhydrogel alone, days 0, 30, sc																
C12 Max	3	3	3	3	3	3	3	5	5	5	3.6	0.966092	0.32203	6		
7 alhydrogel + 27.2 µg F1-WV fusion protein day 0, 30, sc																
CO92 100	28	28	28	28	28	28	28	28	28	28	28	0	0	7		
8 alhydrogel alone, days 0, 30, sc																
CO92 100	3	3	4	4	4	4	6	6	7	7	4.8	1.549193	0.5164	8		
Aerosol:																
9 alhydrogel alone, days 0, 30, sc																
C12 50	3	3	3	3	3	3	3	3	3	3	3	0	0	9		
10 alhydrogel + 13.6 µg F1-WV fusion protein day 0, 30, sc																
C12 50	28	28	28	28	28	28	28	28	28	28	28	0	0	10		
11 alhydrogel + 10 µg Mauro-V urea, days 0, 30, sc																
C12 Max	5	28	7	28	28	28	28	28	28	28	23.6	9.287985	3.096	11		
12 alhydrogel + 13.6 µg F1-WV fusion protein day 0, 30, sc																
C12 Max	28	28	28	28	28	28	28	28	28	28	28	0	0	12		

Challenge group	1	2	3	4	5	6	7	8	9	10	MTD	MTD +/- Stdev	Std error	
13 alhydrogel + 27.2 µg F1-WV fusion protein day 0, 30, sc														
C12 Max	28	28	28	28	28	28	28	28	28	28	28	0	0	13
14 alhydrogel alone, days 0, 30, sc														
C12 Max	3	3	3	3	3	3	3	3	4		3.11111	0.333333	0.11785	14
15 alhydrogel + 13.6 µg F1-WV fusion protein day 0, 30, sc														
CO92 100	28	28	28	28	28	28	28	28	28	28	28	0	0	15
16 alhydrogel alone, days 0, 30, sc														
CO92 100	28	4	3	3	3	3	3	3	3	3	5.6	7.87683	2.62561	16
17 alhydrogel + 10 µg F1 + 10 µg Mauro's V, days 0, 30, sc														
C12 Max	28	28	28	28	28	28	28	28	28	28	28	0	0	17
18 Greer plague vaccine, days 0, 30, sc														
C12 Max	4	4	3	3	3	3	3	3			3.25	0.46291	0.17496	18
19 alhydrogel alone, days 0, 30, sc														
C12 Max	3	3	4	4	5						3.8	0.83666	0.41833	19

HEAD COLOR
WHITE Run 1 AEROSOL
PURPLE Run 2 AEROSOL

34.2, 32.9, 25.7, 35.5, 40.0, 39.0, 37.4, 31.7, 29.2, 41.0 gm = 34.7 avg gm

Prespray -- 4.2×10^9 (48/34/43 on 10^7 plate)

AGC's
#1 -- 2.6×10^7 (18/36/25 on 10^5 plate)

#2 -- 2.2×10^7 (222/196/229 on 10^4 plate)

AEF: 11. EXPOSURE SHEET

Active Immunization

PI: LTC Anderson
Agent: Plague
Strain: C092

Wt: (Ave.) 34.7g

CO92 LD50 = 2.1E + 04

Run #	AGI cfu / ml	AGI cfu	Aerosol cfu / l	MV l	Inhaled Dose cfu	LD50s	Plague Strain
1	2.60E+07	2.60E+08	4.33E+06	0.03	1.30E+06	61.90	C092
2	2.20E+07	2.20E+08	3.67E+06	0.03	1.10E+06	52.38	C092

Protocol: Long Term F1/V

25/26 V/F1 ELISA Summary

Exhibit GA18

Bleed: 21JUN96		Day: Day +204		26JUN96	25JUN96
Plate	Serum	Group	TREATMENT	F1 TITER	V TITER
1A	8906	GP13A	10F1+20V	10,240	20,480
1B	8907	GP13A	10F1+20V	1,280	40,960
2A	8908	GP13A	10F1+20V	10,240	81,920
2B	8909	GP13A	10F1+20V	10,240	81,920
3A	8910	GP13A	10F1+20V	5,120	20,480
3B	8911	GP13A	10F1+20V	20,480	20,480
4A	8912	GP13A	10F1+20V	10,240	40,960
4B	8913	GP13A	10F1+20V	40,960	10,240
5A	8914	GP13A	10F1+20V	2,560	81,920
5B	8915	GP13A	10F1+20V	10,240	81,920
6A	8916	GP13B	10F1+20V	5,120	20,480
6B	8917	GP13B	10F1+20V	20,480	81,920
7A	8918	GP13B	10F1+20V	2,560	40,960
7B	8919	GP13B	10F1+20V	20,480	163,840
8A	8920	GP13B	10F1+20V	10,240	81,920
8B	8921	GP14A	30ugF1-V	2,560	81,920
9A	8922	GP14A	30ugF1-V	2,560	81,920
9B	8923	GP14A	30ugF1-V	1,280	40,960
10A	8924	GP14A	30ugF1-V	1,280	40,960
10B	8925	GP14A	30ugF1-V	1,280	81,920
11A	8926	GP14A	30ugF1-V	2,560	81,920
11B	8927	GP14A	30ugF1-V	5,120	81,920
12A	8928	GP14A	30ugF1-V	1,280	40,960
12B	8929	GP14A	30ugF1-V	2,560	163,840
13A	8930	GP14A	30ugF1-V	5,120	81,920
14A	8931	GP14B	30ugF1-V	10,240	163,840
14B	8932	GP14B	30ugF1-V	20,480	81,920
15A	8933	GP14B	30ugF1-V	10,240	655,360
15B	8934	GP14B	30ugF1-V	5,120	81,920
16A	8935	GP14B	30ugF1-V	2,560	40,960
16B	8936	GP15	PlagueUSP	2,560	2,560
17A	8937	GP15	PlagueUSP	2,560	2,560
18A	8938	GP15	PlagueUSP	2,560	2,560
18B	8939	GP15	PlagueUSP	5,120	2,560
19A	8940	GP15	PlagueUSP	2,560	2,560
19B	8941	GP15	PlagueUSP	5,120	2,560
20A	8942	GP15	PlagueUSP	5,120	2,560
20B	8943	GP15	PlagueUSP	20,480	5,120
21A	8944	GP15	PlagueUSP	40,960	640
21B	8945	GP15	PlagueUSP	5,120	1,280
22A	8946	GP16	ALH alone	640	2,560
22B	8947	GP16	ALH alone	320	640
23A	8948	GP16	ALH alone	320	640
23B	8949	GP16	ALH alone	320	1,280
24A	8950	GP16	ALH alone	320	640
24B	8951	GP16	ALH alone	640	640
25A	8952	GP16	ALH alone	320	1,280
25B	8953	GP16	ALH alone	1,280	2,560
26A	8954	GP16	ALH alone	640	640
26B	8955	GP16	ALH alone	1,280	5,120
27A	F1/V	POOL		327,680	1,310,720
27B	Norm Mouse	POOL		320	2,560

V-titer to high as control serum.

GEOMEAN:	Group	TREATMENT	F1 TITER	V TITER
	GP13A/B	10F1+20V	8,112	44,926
	GP14A/B	30 ugF1-V	3,578	85,794
	GP15	PlagueUSP	5,37	2,229
	GP16	ALH alone	320	1,194

for plague challenge of 5/5/96

170 W.D. 0092
74 LPS 012

data from Ken Walker

JW
8 JUL 96

plague-challenge.sc 7/5/96

7/5/96

P.I. = LTC George Anderson
40 mice, C092 - 100 LD50s
30 mice, C12 - 100 LD50s

Parenteral challenge of mice with C092/M.S. and C12/M.S.

1. Streak 1 slant each with the Master Seed of C092 and C12.
Incubate 2 days at room temperature.
2. Harvest by suspending in 4-5 mls of HIB.
3. Read OD620 of a 1/10 dilution.
4. Adjust to OD 1.0

7/5/96:

Adjusted ODs and read final ODs on 1/2 dilutions:

Final OD = 1.064, for C092

" - 1.10, for C12

C092/M.S.:

1. Prepare dose

5.0 - 7.5 x 10²/ml:

- (1) Add 0.2 ml OD 1.0 to 1.8 mls HIB.
- (2) Add 0.2 ml (1) to 1.8 mls HIB.
- (3) Add 0.5 ml of (2) to 4.5 mls HIB.
- (4) Add 0.5 ml of (3) to 4.5 mls HIB.
- (5) Add 0.5 ml of (4) to 4.5 mls HIB.
- (6) Add 4.0 ml of (5) to 36 mls HIB - - Pipet 10 mls into each of 3 tubes:
mice INOCULUM: 1 x 10³/ml: ~200 cfu/dose

2. Plating: The sample will be diluted in HIB and plated on SBAP:

<u>suspension</u>	<u>Conc./ID</u>	<u>dilution</u>	<u>no. plates</u>	<u>plates</u>
mice inoculum	5 x 10 ² /ml	undil, 10-1	5 each	10

RESULTS:

7/5/96 doses: 1.4 x 10³/ml, 280 cfu/dose (140 LD50s)

7/12/96 doses: 6.5 x 10²/ml, 130 cfu/dose (72 LD50s)

7/18/96 doses: x 10²/ml, cfu/dose (LD50s) - *corrected*

C12/M.S.:

1. Adjust slant suspension to OD620 = 1.0.

Prepare dose

2.3 x 10³/ml:

Manu

34.2

The count

Pres

AGE

#1

#2

- (1) Add 0.2 ml OD 1.0 to 1.8 mls HIB.
- (2) Add 0.2 ml (1) to 1.8 mls HIB.
- (3) Add 0.5 ml of (2) to 4.5 mls HIB.
- (4) Add 0.5 ml of (3) to 4.5 mls HIB.
- (5) Add 1.0 ml of (4) to 9.0 mls HIB.
- (6) Add 6.0 ml of (5) to 18 mls HIB (1/4) - -

INOCULUM, C12-100 sc LD₅₀s (910 cfu). Pipet 10
mls into each of 2 tubes:
1 x 10e3/ml: ~200 cfu/dose

2. Plating: The Inoculum will be diluted in HIB and plated on SBAP:

suspension	Conc./ID	dilution	Total No.	no. plates	plates
C12 Inoculum	2.3x10e3/ml.	undil.		5	
		10-1		5	
		10-2		5	
				TOTAL-15	

RESULTS:

7/5/96 doses:

7/12/96 doses:

7/18/96 doses:

3.36 x 10e3/ml. 6.7 x 10e2 cfu/dose (73.6 LD50s)
3.0 x 10e3/ml. 5.8 x 10e2 cfu/dose (63.7 LD50s)
x 10e3/ml. x 10e2 cfu/dose (LD50s) - *corrected*

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**AIBS PEER REVIEW TO USAMRMC
MEDICAL BIOLOGICAL DEFENSE RESEARCH PROGRAM
ON PLAGUE**

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KATHLEEN McDONOUGH, P.A.D.

DATE: March 13, 1996

INTRODUCTION

AIBS was requested by US Army Medical Research and Development Command (USAMRDC) to convene a review Panel to provide an assessment of the scientific merit of the Medical Biological Defense Research Program (MBDRP) on Plague. It was requested that the three scientific reviewers have a collective knowledge of the following subject areas: *Yersinia pestis*, Vaccine Production, Molecular Genetics and FDA requirements for a vaccine. Such a panel was convened and provided with documentation by USAMRDC to read prior to the conference. This consisted of abstracts prepared by the individual investigators who form the MBDRP on Plague (see Appendix 1.)

CHARGE TO PANEL

Three scientific reviewers were asked to evaluate the MBDRP on Plague. They independently reviewed material provided by USAMRDC and attended a conference on the subject matter. They were asked to judge the scientific merits of the Program.

The reviewers, individually, provided comments to AIBS, who in turn compiled this written report summarizing these comments and the discussions at the conference. The Chairman of the Review Panel read and approved the report prior to its submission to USAMRDC.

PRESENTATION SUMMARIES

The conference comprised presentations by each of the following investigators. Abstracts were provided for by each and are attached as Appendix 1.

COL ARTHUR FRIEDLANDER

Overview of plague program

COL RUSSELL BYRNE

Antibiotic treatment of experimental pneumonic plague

DR. PATRICIA WORSHAM, DR. M. LOUISE PITT, LTC KELLY DAVIS

F1 is not a required virulence factor for the mouse or non-human primate

MAJ GERALD P. ANDREWS, LTC GEORGE J. ANDERSON, JR.

Protective efficacy of active immunization with purified F1 from *Yersinia pestis* and an *Escherichia coli* recombinant strain against lethal parenteral and respiratory plague challenge

DR. PATRICIA WORSHAM

Studies on the role of the pigmentation locus in the pathogenesis of *Y. pestis*

DR. SUSAN L. WELKOS, LTC KELLY J. DAVIS

Analysis of the role of pPst encoded genes in pathogenesis of infection by *Y. pestis*

DR. ALAN SAMPLE

Plasminogen activator protease degrades proinflammatory cytokines

MAJ GERALD P. ANDREWS, DR. SUSAN STRALEY, DR. ALAN SAMPLE, MAJ

GERALD P. ANDREWS

Cloning, Expression, Purification, and Protective Efficacy of Yops and pH 6 antigen

LTC GEORGE J. ANDERSON, JR., DR. DAVID HEATH

Cloning, expression, and protective efficacy of V antigen

LTC GEORGE J. ANDERSON, JR.

Cloning, expression, and protective efficacy of F1-V fusion protein

DR. JEFFREY PULLEN

Determination of important B and T-lymphocyte epitopes in the F1 and V antigen proteins of *Yersinia pestis*

COL ARTHUR FRIEDLANDER

Overview of future plans

SUMMARY EVALUATIONS OF THE RESEARCH AREAS

The review panel read the abstracts provided by the investigators prior to the meeting on February 15, 1996, and listened to presentations by each of the investigators at the meeting. The following comments include recommendations to individual investigators, and are intended to be constructive. Certain points apply to more than one project, or even to the program as a whole, and hence may appear repetitive. Also, the reviewers recognize that some of their recommendations may be affected by programmatic decisions that are beyond the control of the immediate Program staff and thus may not prove to be possible.

COL ARTHUR FRIEDLANDER

Overview of plague program

The USAMRMC Plague Research Program's primary objective is to develop a vaccine that will protect military personnel if exposed to an aerosol attack of *Yersinia pestis*, the causative agent of plague. Given that the currently available vaccine (USP) protects primarily through anti-F1 antibody, that this vaccine offers very poor protection from primary pneumonic plague, and that F1⁻ strains are highly virulent, there clearly is a need for a new, more protective vaccine. Once developed, the general population living in areas endemic for plague would also benefit from such a vaccine.

Most of the projects presented as separate studies and presentations clearly meet the program's primary objective. Part of the rationale for the approach taken is that an aerosol attack of *Yersinia pestis* might include strains that do not produce the F1 capsular antigen. Given that the current vaccine (USP) stimulates primarily antibodies

**AIBS PEER REVIEW TO USAMRDC
MEDICAL BIOLOGICAL DEFENSE RESEARCH PROGRAM
ON PLAGUE**

TIME: February 15, 1996, 8.00am to 5.00 pm

LOCATION: US Army Medical Research Institute of Infectious Diseases,
Conference Room, Fort Detrick, Frederick, MD

EXECUTIVE SUMMARY

Overall, the program has made very significant and impressive advances in only a few years towards the development of a new vaccine, and Dr. Friedlander and his entire team of investigators can be proud of their accomplishments to date. They clearly have a very viable, sound program with a good team of investigators that is focused with high potential to succeed. It is hoped that the administration will continue to support this effort and provide the group with the resources and time necessary to complete their task. The investigators clearly considered the recommendations of the previous reviewers and incorporated several of the suggestions into their program.

The team has invested significant effort in examining numerous virulence determinants of *Yersinia pestis* for their ability to stimulate protection through immunization. The F1 capsular antigen and the V antigen have been shown by investigators in other laboratories to be good candidates for inclusion in a new multivalent subunit vaccine. The team at USAMRMC has confirmed the protective value of these two antigens. However, realizing that F1 and V antigens might not be sufficient for full protection against all virulent strains of *Y. pestis*, the group has worked through an impressive list of additional candidates. The only other antigen that offered significant protection was YopD, although protection was only observed when mice were challenged with the F1⁻ strain. Passive immunizations with anti-F1 and anti-YopM antisera deserve further attention. Combined antibiotic treatment and immunization might increase the survival of animals challenged by aerosol.

The team appears to make use of mice and nonhuman primates as excellent animal models for both their parenteral and aerosol challenge experiments. The current vaccine study protocols for test challenges are very good.

The development of *in vitro* correlates of immunity should be a high priority of the program. It is currently the weakest portion of the future plans. As discussed with the investigators, the assumption that protection is solely antibody-mediated has potential difficulties. Before continuing studies to map active B cell epitopes, the investigators need to determine the role of T cells in immunity to plague.

examined for *Y. pestis* in the LD50 studies and survivors were examined for clearance of the organisms to determine the full level of protection provided by vaccination.

In the first study, the V antigen was examined for its ability to generate a protective immune response in mice challenged by parenteral subcutaneous or aerosol challenge with either the F1⁺ or F1⁻ isogenic strains of *Y. pestis*. Recombinant V antigen was cloned and expressed in two fusion/expression systems and used with an adjuvant approved for human use (Alhydrogel). Both preparations of rV antigen were administered twice and provided very good protection in mice challenged by both routes and both strains. This is an excellent study and identifies (as another laboratory has demonstrated independently) the V antigen as an excellent candidate immunogen to include in a vaccine to protect from aerosol infections with either F1⁺ or F1⁻ strains. These studies are critical to the program's objective and provides some quite exciting results.

The second study extends the work on the V antigen of *Y. pestis* by examining protection following a single dose of 10 µg (the previous study used two immunizations prior to challenge). Mice were subsequently challenged by aerosol exposure to either low and high doses of the F1⁺ or F1⁻ strain. Protection ranged from 70% to 78% survival in these mice, demonstrating that a single immunization could afford significant protection from an aerosol route of infection. However, the schedule including a primary immunization followed by a single boost afforded 20% to 30% greater protection (previous report). While it is of interest what level of protection results from a single dose, future work with nonhuman primates will likely confirm what we know about many other bacterial vaccines, i.e., better protection results with boosts following the primary immunization.

Two areas need to be addressed in future work on the V antigen. The studies presented used the V antigen tagged with histidine from the pET vector. If this antigen is to be used in humans, a method for the efficient removal of the his-tag is needed. Identifying the active sites on the V antigen responsible for protective immunity as well as potential negative biological activities, such as immune suppression, may be required for this antigen to be safe. The group might also consider examining how long protective immunity lasts following vaccination with the V antigen. Some of these issues were addressed by Dr. Friedlander in his closing remarks.

LTC GEORGE J. ANDERSON, JR.

Cloning, expression, and protective efficacy of F1-V fusion protein (abstract 17)

Prior studies have confirmed the potential for both F1 and V antigen to protect mice from *Y. pestis* by both parenteral and aerosol routes. In this study a construct was made containing the F1 and V antigen genes for expression of a fusion protein. When the F1-V fusion protein was used for immunization, mice were protected when challenged by needle or aerosol with either the F1 positive or F1 negative strain of *Y. pestis*. Poorer protection resulted when only a portion of the V antigen was expressed as a fusion protein with F1. This work is quite clever and interesting, and advances the program's effort towards the development of a multivalent vaccine. The attempt to make fusions of these two antigens also demonstrates an advance towards reducing

the steps required for making and purifying antigens for the vaccine. The investigators are also testing longer term antibody responses and how long protection lasts (a concern raised from the previous studies with the V antigen alone). Antibody responses to the F1 and V antigen components of the fusion protein were also examined. Both F1 and the V antigen have been shown by other workers to be protective and now the group at USAMRMC has shown that rF1 and rV are the best candidates identified to date for a new plague vaccine.

Again, this fusion protein has a histidine tag, which will need to be removed prior to its use in humans.

DR. JEFFREY PULLEN

Determination of important B and T-lymphocyte epitopes in the F1 and V antigen proteins of *Yersinia pestis* (abstract 18)

This study attempts to identify important B and T cell epitopes within both the F1 and V antigens, however only B cells were addressed in the presentation. Identifying the functional epitopes in these proteins is important both to an understanding of the protective mechanisms stimulated by these two immunogens, and for assessing the potential of using synthetic peptides rather than entire recombinant proteins in a vaccine. This study is an important part of fulfilling the long-term objective of developing a useful vaccine. However, the usefulness of the current approach should be carefully reconsidered.

The use of short peptides to generate antibodies without conjugation to carrier molecules has, in general, not been very successful. Although it is sometimes possible to generate antibodies against short peptides, it is unlikely that the response will be protective without some T cell involvement. The investigators' initial experiments showed that peptides generated from the region of the protein known to be antigenic failed to generate a protective response despite generating significant antibody production. These results should have alerted them to the problems inherent in this approach. Instead, the investigators expanded their studies in response to these findings by making and testing additional peptides covering the whole of V antigen and F1 protein. This was a lot of work, using a lot of mice, that generated very little useful information. A simpler and more direct approach to begin mapping the reactive epitopes in these immunogens is to screen the overlapping peptides *in vitro* using antisera from animals or humans that have either had infections with *Y. pestis* or been immunized with native F1 and/or V antigen. Another concern is that in the future goals, it was stated that the response to the peptides, rather than to the native antigen will be tested to better determine the response. However, since the goal is to get protective antibodies, it seems that the response to native antigen, which is what the animals will see in an actual infection, is what should be measured.

It is also important for the investigators to determine the nature of a protective immune response to *Y. pestis* infection before restricting their focus and undertaking such labor-intensive studies to define only B cell epitopes. Antibody reactivity does not assure protection, and with some pathogens high antibody titers have even been correlated with disease progression. In addition, non-F1 antigens may evoke a

COMBINED RECOMMENDATIONS AND CONCLUSIONS

The USAMRMC's program to develop a new subunit vaccine for pneumonic plague has been very productive and has made significant advances towards this objective. The leader and research team are highly skilled, competent investigators and, with continued support, it is anticipated that a new vaccine for human trials is only a few years away. The investigators have used very effective immunization and challenge protocols to test immunogens in both mice and nonhuman primates for protection against plague following either parenteral or aerosol exposures to *Yersinia pestis*. Having the facilities to safely execute aerosol transmission studies is a critical component of this program. The team has confirmed and extended the data supporting the potential for both recombinant F1 and V antigens to afford significant protection. The work using the F1-V antigen fusion protein is exciting and represents a significant advance made by this team.

The team has examined numerous other antigens for identifying additional protective immunogens, especially for challenge with strains of *Y. pestis* lacking the F1 antigen. For such isolates, the V antigen and possibly YopD are the only useful candidates identified to date. The addition of one more antigen would likely solve the problem of non-responders, as well as strengthen the response in all individuals. The choice of antigens being tested for potential vaccine components appears somewhat random. These studies could be focused better by determining what proteins induce an immune response, thereby demonstrating which determinants are most likely being seen by the immune system. Although it is not possible to predict in advance which antigens are protective, the search could have been directed more towards antigens known to induce an antibody response in infected human patients and laboratory infected animals. Additional focus on the basis of immunity to plague challenge is also recommended. The investigators are also aware of the immunosuppressive effects of V antigen, and plan to examine the mechanisms involved. These types of studies should allow the team to "fine tune" the V antigen to increase its efficacy and safety as a vaccine component.

The development of *in vitro* correlates of immunity should be a high priority of the program and is currently the weakest area of the future plans. As discussed with the investigators, the assumption that protection is solely mediated by antibody has potential difficulties. Before continuing studies to determine important B cell epitopes, the role of T cells needs to be addressed in collaboration with immunologists. There are standard methods, such as adoptive transfer, to determine if T cells protect against challenge with *Y. pestis*. There are also *in vitro* techniques to determine if T cells taken from an immunized animal proliferate in response to specific antigens. The studies using synthetic peptides have potential, but this work needs to be done with conjugated peptides. Alternatively, peptides could be attached to larger inert particles that could be taken up by B cells or macrophages that then present the antigen on class II MHC molecules on their surface. Epitope mapping of the F1 and V antigen peptides using immune sera from natural infections would have been an appropriate first step.

APPENDICES

APPENDIX 1: AGENDA

APPENDIX 2: ABSTRACTS

REVIEW OF PLAGUE RESEARCH PROGRAM

USAMRIID

15 FEBRUARY 1996

0815-0830 Welcome and introduction
COL David Franz, DVM, Ph.D.

0830-0900 Overview of Plague Program
COL Arthur M. Friedlander, M.D.

Treatment

0900-0930 Antibiotic treatment of experimental pneumonic plague
COL Russell Byrne, M.D.

Role of F1 Capsule in Pathogenesis and Immunity

0930-1000 Protective efficacy of active immunization with purified F1
from *Yersinia pestis* and an *Escherichia coli*
recombinant strain against lethal parenteral and respiratory
plague challenge
MAJ Gerard P. Andrews, Ph.D.

1000-1015 Coffee Break

1015-1100 F1 capsule is not a required virulence factor for the mouse or
non-human primate
Patricia L. Worsham, Ph.D.
M. Louise Pitt, Ph.D.
LTC Kelly J. Davis, DVM

Role of Non-F1 Proteins in Pathogenesis and Immunity

1100-1130 Studies on the role of the pigmentation locus in the
pathogenesis of *Y. pestis*
Patricia L. Worsham, Ph.D.

1130-1300 Lunch

- 1300-1320 Analysis of the role of pPst encoded genes in pathogenesis of infection by *Y. pestis*
Susan L. Welkos, Ph.D.
- 1320-1335 Plasminogen activator protease degrades proinflammatory cytokines
Allen Sample, Ph.D.
- 1335-1405 Cloning, expression, and protective efficacy of Yops and pH 6 antigen
MAJ Gerard Andrews, Ph.D.
- 1405-1420 Cloning, expression, and protective efficacy of V antigen
LTC George J. Anderson, Jr., Ph.D.
- 1420-1435 Cloning, expression, and protective efficacy of F1-V fusion protein
LTC George J. Anderson, Jr., Ph.D.
- 1435-1450 Determination of important B and T-lymphocyte epitopes in the F1 and V antigen proteins of *Y. pestis*
Jeffrey Pullen, Ph.D.
- 1450-1515 Overview of future plans
COL Arthur M. Friedlander, M.D.

Recombinant F1-V (rF1-V) Fusion Protein Protects against Lethal
Wildtype *Yersinia pestis* in a Mouse Model

DAVID G. HEATH, GEORGE W. ANDERSON, JR., CHRISTOPHER BOLT, SUSAN
L. WELKOS, PATRICIA L. WORSHAM, AND ARTHUR M. FRIEDLANDER
Bacteriology Division, U.S. Army Medical Research Institute of
Infectious Diseases, Ft. Detrick, MD.

The virulence of F1- strains and their occurrence in nature imply that F1 immunogen will not be sufficient for an optimal new plague vaccine. A fusion protein has the theoretical possibility of simplifying and reducing the cost of production of multiple antigens in addition to stabilizing the protein. For these reasons, we developed a fusion protein consisting of both the F1 and V antigens (1). The first fusion protein made consisted of F1 fused with residues 168-175 of the V antigen, a segment which previous studies suggested to contain a protective epitope. This fusion protein was used with the adjuvant alhydrogel (aluminum hydroxide) to immunize female Swiss Webster (Hsd:ND4) mice subcutaneously (s.c.) on days 0 and 30 followed by a s.c. or aerosol challenge with either the F1- C12 strain (LD50 = 9.1 CFU, s.c.; LD50 = 1.1×10^5 CFU, aerosol route) or the wild-type F1+ CO92 (LD50 = 1.9 CFU, s.c. route; 2.1×10^4 CFU, aerosol route) strain of *Y. pestis*. Endotoxin had been removed from the preparations prior to immunization, so that this would not be a confounding factor.

When 18.5 μ g of the F1-V168-275 fusion protein was used to immunize mice, there was 90% survival (9/10) when challenged s.c. with 63 LD50 of the F1+ CO92 strain. The positive control was a group of mice immunized with 10 μ g of rF1 which is equivalent to the F1 content of the F1-V168-275 protein. The rF1 control resulted in 100% (10/10) protection. The F1-ELISA IgG titers were the same (1:81920). All mice in alhydrogel control group died (0/9; MTD \pm SD, 5.2 ± 1.0). When the F1-V168-275 immunized mice were challenged with 104 LD50 by the aerosol route, protection was 80% (8/10; MTD \pm SD, 20.3 ± 7.1) compared to 0% for the control group (0/10; MTD \pm SD, 3.1 ± 0.3 ; 80-104 LD50). The group immunized with rF1 resulted in 70% protection (7/10; MTD \pm SD, 9.0 ± 1.0) when challenged with 80 LD50. The addition of part of the V protein onto the F1 protein did not appear to effect its antigenicity.

The F1- strain, C12, was used to test the ability of the partial V portion of the F1-V168-275 protein to protect mice against a lethal challenge. Here 27 μ g of the F1-V168-275 fusion protein was used, which is equivalent to 10 μ g of the V protein

known to be protective. A s.c. challenge dose of 55 LD50 resulted in 30% survival (3/10, MTD \pm SD, 9.4 ± 7.0). All of the controls died (0/10, MTD \pm SD, 10.8 ± 4.8). While there was some protection, there was no increase in the MTD. There was a good V-ELISA antibody response to the F1-V168-275 (1:163840). In case this response was not sufficient, another group was immunized with 27 μ g, but with complete Freund's adjuvant (CFA). In this case, protection was only 20% (2/10, MTD \pm SD, 9.1 ± 3.2), while 10 μ g of rV in CFA resulted in 100% protection. The V-ELISA titer when CFA was used was 1:1310720 for F1-V168-275 and rV. A 10-fold increase in the V-antibody titer did not have any effect on protection and the V-ELISA titer was not indicative of protection. When a group of F1-V168-275 mice were challenged with 95 LD50, C12, by the aerosol route, no mice survived (0/10, MTD \pm SD, 3.5 ± 0.5). All of the alhydrogel control group died (0/10, MTD \pm SD, 3.4 ± 0.5). In other experiments, rV itself gave 80-90% protection against an aerosol challenge.

These results demonstrated the feasibility of making a F1-V fusion protein. The efficacy of F1 was not altered by making a fusion protein. However, while the V168-275 protein portion of the fusion protein was antigenic, it was not immunogenic. This caused us to address the question as to whether the entire V protein could be fused to F1 and whether it would be immunogenic.

Using a fusion protein which combines the whole F1 and the whole V protein (rF1-V) to immunize mice on days 0 and 30 increased the protection afforded by the V portion of the fusion protein. When 13.6 μ g of rF1-V was used to immunize mice, there was 100% (10/10) protection against a s.c. challenge of 57 LD50 and 90% (9/10) protection against 1.1×10^6 LD50 C12 strain. Ten micrograms (10 μ g) of rV also gave 90% (9/10) protection against 1.1×10^6 LD50, C12 strain. All of the alhydrogel control group died (0/10, MTD \pm SD, 6.0 ± 0.0). The rF1-V protein also offered protection against an aerosol challenge. The same immunization schedule resulted in 100% (10/10) when mice were challenge with 546-636 LD50, C12 strain on day 73 postimmunization. When mice immunized with the rF1-V fusion protein were challenged with 762 LD50 of the F1+, CO92 strain by the aerosol route, 100% (10/10) of the mice survived. The F1-V fusion protein was able to protect mice from a significant aerosol challenge from either a F1+ or F1-lethal strain of *Y. pestis*. This protection is better than the protection afforded by the current Plague Vaccine USP. When mice which were immunized on day 0 and 30 with 0.2 ml of the current vaccine and challenge by the aerosol route on day 73 postimmunization with 546-636 LD50, C12 strain, all of the mice died (0/8, MTD \pm SD, 3.3 ± 0.5). The V-ELISA titer to the Plague

Vaccine USP was <1:640.

The recombinant rF1-V fusion protein was produced in *E. coli* and contained a polyhistidine tag which aids in the purification of the fusion protein. While this protein has been shown to be highly efficacious in the mouse model, it remains to be seen whether this level of protection will be seen in the non-human primate model. Further, the regulatory issue of whether a histidine tagged protein will be acceptable to the Food and Drug Administration needs to be resolved.

1. Heath, D.G., G.W. Anderson, Jr., J. M. Mauro, S.L. Welkos, and A.M. Friedlander. Protection against experimental bubonic and pneumonic plague by a recombinant capsular F1-V antigen fusion protein vaccine. manuscript submitted.
2. Brubaker, R.R., A.K. Sample, D.Z. Yu, R.J. Zahorchak, P. C. Hu, and J.M. Fowler. 1987. Proteolysis of V antigen from *Yersinia pestis*. *Microbial Pathogenesis*. 2:49-62.

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